

State of Hawaii
Department of Agriculture
Plant Industry Division
Plant Quarantine Branch
Honolulu, Hawaii

June 28, 2022

Board of Agriculture
Honolulu, Hawaii

Subject: (1) Provided the Southern House Mosquito, *Culex quinquefasciatus*, is Placed on the List of Restricted Animals, Part A, Allow the Importation of Lab-Reared Strains of the Mosquito, *Culex quinquefasciatus* (Diptera: Culicidae), Inoculated with Strains of *Wolbachia* Bacteria, by Permit, For Immediate Field Release to Suppress Wild Populations of *Culex quinquefasciatus*, by the Hawaii Department of Land and Natural Resources; and

(2) Provided the Southern House Mosquito, *Culex quinquefasciatus*, is Placed on the List of Restricted Animals, Part A, Establish Permit Conditions for the Importation and Immediate Field Release of Lab-Reared Strains of the Mosquito, *Culex quinquefasciatus* (Diptera: Culicidae), Inoculated with Strains of *Wolbachia* Bacteria to Suppress Wild Populations of *Culex quinquefasciatus*, by the Hawaii Department of Land and Natural Resources.

I. Summary Description of the Request

PQB NOTES: *The Plant Quarantine Branch (PQB) submittal for requests for import or possession permits, as revised, distinguishes information provided by the applicant, Suzanne Case, from procedural information and advisory comment and evaluation presented by PQB. With the exception of PQB notes, hereafter "PQB NOTES," the text shown below in section III from page 4 through page 18 of the submittal was taken directly from the applicant's application and subsequent written communications provided by the applicant. For instance, the statements on pages 13 through 15 regarding effects on the environment are the applicant's statements in response to standard PQB questions and are not PQB's statements. This approach for PQB submittals aims for greater applicant participation in presenting import requests in order to move these requests to the Board of Agriculture (Board) more quickly, while distinguishing applicant provided information from PQB information. The portion of the submittal prepared by PQB, including the procedural background, summary of proposed list additions, environmental assessment, proposed permit conditions and advisory*



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Culex quinquefasciatus
Field Release
Suzanne Case, DLNR

Board of Agriculture

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review, are identified as sections II, IV, V, and VI of the submittal, which start at pages 3, 18, 19, and 24 respectively.

COMMODITY: Various Shipments of the Southern House Mosquito, *Culex quinquefasciatus* (Diptera: Culicidae), inoculated with Strains of *Wolbachia* Bacteria.

SHIPPERS: Stephen Dobson
MosquitoMate, Inc.
2520 Regency Road,
Lexington, Kentucky, 40503

Verily Life Sciences
269 E Grand Avenue,
South San Francisco, California 94080

IMPORTER: Suzanne Case, Chairperson
Hawaii Department of Land and Natural Resources
1151 Punchbowl Street, Honolulu, HI 96813

CATEGORY: *Culex quinquefasciatus* is currently an unlisted animal. Animals not found on any list are considered prohibited until placed on a list. Additionally, Chapter 4-71, Hawaii Administrative Rules (HAR), allows importation of unlisted animals into Hawaii under special permit for the purpose of remediating medical emergencies or ecological disasters, or conducting scientific research that is not detrimental to agriculture, the environment, or humans by special permit, on a case-by-case basis, as approved by the Board.

II. Procedural Background

DLNR has requested that one of the lists in Chapter 4-71, Hawaii Administrative Rules (HAR), be amended by Board Order to include the Southern House Mosquito, *Culex quinquefasciatus*. The species may be placed on the List of Conditionally Approved Animals, List of Restricted Animals (Part A or B), or the Prohibited List. Species on the Restricted and Conditionally Approved Lists may enter the State of Hawaii under permits with conditions approved by the Board. Until placement on a list, species are considered prohibited except as provided by Section 150A-6.2(c), Hawaii Revised Statutes (HRS).

Species on the List of Restricted Animals (Part A) are available for research by universities and government agencies, exhibition in municipal zoos and government-affiliated aquariums, and for other institutions for medical and scientific purposes as determined by the Board. All species listed for import require a permit for entry into the State.

Pursuant to HRS §150A-6.6, the Board has the authority to adopt administrative rules to make additions to or deletions from the lists required to be maintained under HRS §150A-6.1 through §150A-6.3, which include the List of Restricted Animals, Part A. Changes to the lists can be made without regard to the notice and public hearing requirements of HRS Chapter 91 provided that there is notice and opportunity for public input regarding additions or deletions to the lists.

HAR §4-71-4.2, "Public Input and Notification for Listing," details the specific process that the Board must follow to make a change to the lists maintained by PQB. It requires that, thirty days or more prior to the effective date of the Board order, the Hawaii Department of Agriculture (Department) issue a press release and mail a notice to the Office of Environmental Quality Control, now the Environmental Review Program, for publication and to all persons who have made a timely written request of the department for advance notice of the order or the Department's rulemaking proceedings.

Provided the Board acts favorably on this request for list placement by Board Order, the species will have been placed on a respective list and be eligible for import and/or possession. PQB can then process a permit application by having the Board approve the future importation and establishment of appropriate permit conditions for the organism and proposed purpose.

III. Information Provided by the Applicant in Support of the Application

Summary Description of the Requests

In accordance with the provisions of Chapter 150A, Hawaii Revised Statutes, we are requesting to import the following animal commodities:

Commodity	Scientific Name	Quantity
Southern House Mosquitoes (Male Adults)	<i>Culex quinquefasciatus</i>	Continued shipments for immediate release.

Additionally, we are requesting the listing of *Culex quinquefasciatus* mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

Reason for importation:

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Culex quinquefasciatus
Field Release
Suzanne Case, DLNR

Board of Agriculture

For immediate field release applications to suppress mosquito populations in areas where Hawaii fauna are at risk of disease transmission due to the presence of these mosquitoes.

Shippers:

- 1) Stephen Dobson, MosquitoMate, Inc.
2520 Regency Rd., Lexington, KY, 40503
- 2) Verily Life Sciences
269 E Grand Ave, South San Francisco, CA 94080

Importers:

- 1) DLNR Waimano Baseyard – Hawaii Invertebrate Program - Oahu
2680 Waimano Home Road, Pearl City, HI 96782, (808) 266-7989
- 2) Kaua'i Branch, Division of Forestry & Wildlife, 3060 Eiwa Street Rm. 306, Lihue, HI 96766. (808) 274-3433
- 3) O'ahu Branch, Division of Forestry & Wildlife, 2135 Makiki Heights Drive, Honolulu, HI 96822. (808) 973-9778
- 4) Maui (& Moloka'i) Branch, Division of Forestry & Wildlife, 1955 Main Street, Room 301, Wailuku, HI 96793. (808) 984-8100
- 5) Hawai'i Branch, Division of Forestry & Wildlife, 19 E. Kawili Street, Hilo, HI 96720. (808) 974-4221

Project:

This is an application for:

- A permit to import male *Culex quinquefasciatus* mosquito species.
- The listing of *Culex quinquefasciatus* mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

As outlined in the suggested conditions for importation, these mosquitoes will either contain the same wild type bacterium (*Wolbachia* spp.) which is already endemic in *Culex quinquefasciatus* mosquitoes in Hawaii, or will be inoculated with an incompatible bacterium (*Wolbachia* spp.) that is not native to the wild mosquito's current internal fauna. The presence of this different strain of bacteria within the male mosquito's reproductive system will render the imported male mosquitoes unable to successfully mate with wild females found within Hawaii, a process called cytoplasmic incompatibility. Cytoplasmic incompatibility has been used with much success in other parts of the world to reduce mosquito populations and thus reduce the potential of transmission of mosquito vectored diseases. We intend to import male, sexually

incompatible mosquitoes for direct release onto the environment. This process uses cytoplasmic incompatibility to reduce current populations of this pest mosquito species, which are vectors for pathogens to Hawaii's fauna, including pathogens such as avian malaria, and which can vector West Nile virus, and lymphatic filariasis to humans. Importing Hawaii lineage mosquitoes which contain the wild type bacterium, will ensure that we can conduct genetic analysis to confirm that the wild *Culex quinquefasciatus* is the wild type originally provided to the collaborators, and that the inoculated mosquitoes are indeed incompatible.

Culex quinquefasciatus is an invasive, disease-spreading mosquito that has dispersed across the Hawaiian islands since its accidental introduction in the 1800s. The species is present on Hawaii, Maui, Molokai, Lanai, Kahoolawe, Oahu, Kauai, and the northwest Hawaiian islands. *Culex quinquefasciatus* can thrive at sea-level to 4800ft in elevation. In Hawaii, the mosquito is able to transmit pathogens to native forest birds. The spread of avian malaria, in particular, has contributed to the extinction of more than half of Hawaii's endemic honeycreepers and continues to pose a risk to the remaining species. *Culex quinquefasciatus* is also known to transmit dog heartworm within pets found throughout Hawaii, and is a concern to human health given its ability to vector West Nile virus on the US mainland and lymphatic filariasis in other Pacific nations.

Efforts to suppress *Culex quinquefasciatus* through utilization of traditional vector

control methods (e.g., pesticides) are inadequate at a landscape scale, and may be problematic for other non-target state and federally protected invertebrate species including Hawaiian picture-wing flies (*Drosophila* spp.), damselflies (*Megalagrion* spp.), yellow-faced bees (*Hylaeus* spp.) and anchialine pond shrimps (*Vetericaris chaceorum* and *Procaris hawaiiana*). Current efforts to control mosquito-vector disease outbreaks are limited to reducing mosquito breeding site locations and localized applications of various larvicides and adulticides.

On September 6-7, 2016, local, national, and international experts gathered in Hawaii to discuss how to mitigate mosquito-borne diseases. The strategy deemed most favorable in terms of its effectiveness, technical readiness, and safety was *Wolbachia*-based cytoplasmic incompatibility. Cytoplasmic incompatibility results from the presence of a bacterium, *Wolbachia*, in the cells of the mosquito. Many arthropod species, including several native species here in Hawaii, naturally contain strains of *Wolbachia*. Bacteria in the genus *Wolbachia* are a type of arthropod endosymbiont that do not occur in humans or other vertebrates. Approximately 50% of insect species naturally have the bacteria, although many of these insects can survive without *Wolbachia*. Conversely, *Wolbachia* cannot persist outside of insect cells, as it is an obligate endosymbiont. The largest effect of *Wolbachia* is on mating compatibility between individual insects that carry the bacteria. However, there are secondary effects that are being studied by many labs. These include altered host insect lifespan and reduced vector competence.

In nature, *Wolbachia* are passed from females to their offspring. Different strains of

Wolbachia have also been introduced into insects in laboratories. If a male mosquito with one type of *Wolbachia* mates with a female mosquito that has a different strain of *Wolbachia* the resulting offspring can be inviable and not develop into mosquito larvae because of a mismatch of cellular signals (loss of the male parental chromosomes) originating from *Wolbachia*. If sufficient numbers, on the order to 10 times the wild population size, of male mosquitoes of a different *Wolbachia* type are released, wild females are more likely to mate with males of a different *Wolbachia* type and are predicted to have far fewer viable offspring. With subsequent releases, this process can significantly suppress the wild population numbers of mosquitoes over the following generations over a geographic area. *Wolbachia* male-based insect control programs have been highly successful for reducing local mosquito populations around the world. Results of initial trials in Fresno, California showed decrease of biting *Ae. aegypti* females by 68%, 95%, and 84% during the peak mosquito seasons in 2017, 2018, and 2019 respectively. *Wolbachia* cannot be spread by the released males, because *Wolbachia* are only passed from mother to offspring. It is also worth noting that male mosquitoes do not bite or vector disease.

One way to generate mosquitoes with a different *Wolbachia* type, is by clearing the naturally-occurring *Wolbachia* strain from the mosquitoes using the antibiotic tetracycline. Then *Wolbachia* can be harvested from cells of another insect species (this can be another mosquito or a non-mosquito species) and introduced into the cleared

mosquitoes via microinjection. Another method to establish new *Wolbachia* strains is to mate a *Wolbachia*-carrying female insect to males that have been cleared of their naturally-occurring *Wolbachia* via antibiotic treatment. Because *Wolbachia* are maternally inherited (described above), this cross results in all of the offspring inheriting whichever *Wolbachia* strain is contained in the female parent. Incompatible *Wolbachia* strains can also be naturally present in populations of mosquitoes.

The first shipper listed within this import application, MosquitoMate Inc., holds the US patent, Patent No.: US 7,868,222 B1, for the method of producing an artificial infection in Culicidae species.

(<https://patentimages.storage.googleapis.com/55/da/ae/d7cb8b9cb44599/US7868222.pdf>)

Additionally, MosquitoMate Inc. offers a commercially available, *Wolbachia* infected male mosquito product for purchase to suppress *Aedes albopictus* mosquito populations via cytoplasmic incompatibility. This product, ZAP Males®, has been reviewed and registered under the Federal Insecticide, Fungicide, and Rodenticide Act (FIFRA). ZAP Males® are a labeled pesticide product with the EPA registration number 89668-4. This product currently has a restriction that only allows for its application in specific states, which does not currently include Hawaii. We reference this, as it is the only registrant in the US currently with a *Wolbachia* mosquito product currently in process of registration with the EPA.

(https://www3.epa.gov/pesticides/chem_search/ppls/089668-00004-20171103.pdf)

The second shipper listed is Verily Life Sciences, a CA based company which is in the process of working with a different incompatible *Culex quinquefasciatus*. This company is initiating consultations with the EPA relating to this different *Wolbachia* mosquito and will provide additional information directly to HDOA as needed.

Culex quinquefasciatus mosquito eggs originating from Hawaii stock (aka collected from field sites in Hawaii) have been provided to MosquitoMate and Verily for development and testing of cytoplasmic incompatibility. These mosquitoes have been crossed with female mosquitoes carrying a different *Wolbachia* species as outlined above. These mosquitoes have then been backcrossed with a separate population of mosquitoes originating from Hawaii stock over at least seven generations to ensure Hawaii's wild mosquito genetics are >99% contained within a commercially available product to be applied within Hawaii.

Generations	HI Mosquito Genetics	Crossed MosquitoMate Genetics
0	100.00%	100.00%
1	50.00%	50.00%
2	75.00%	25.00%
3	87.50%	12.50%
4	93.75%	6.25%
5	96.88%	3.13%
6	98.44%	1.56%
7	99.22%	0.78%
8	99.61%	0.39%
9	99.80%	0.20%
10	99.90%	0.10%

On January 17, 2017, the Hawaii Invasive Species Council, an inter-departmental collaboration of the Departments of Land and Natural Resources (DLNR), Agriculture (HDOA), Health (HDOH), Transportation (DOT), Business, Economic Development & Tourism (DBEDT), and the University of Hawaii (UH) passed resolution 17-2, specifically pertaining to mosquitoes. Resolution 17-2, entitled, "Supporting Evaluation and Implementation of Technologies For Landscape-Scale Control of Mosquitoes, With a Focus On Mitigating Both Human and Wildlife Health Risks," recognizes that mosquitoes in the State of Hawaii are non-native and an important pest species to control. The resolution supports the implementation of evaluated technologies that are

scientifically demonstrated as safe, effective control measures for mosquitoes.
(<https://dlnr.hawaii.gov/hisc/files/2013/02/HISC-Reso-17-2-signed.pdf>).

House Resolution (HR) 297 passed the Hawaii State House in 2019 and further directed “DOA to review the *Aedes aegypti* mosquito with *Wolbachia* bacteria, including *Aedes aegypti* mosquitoes originating from Hawaii stock that could be imported for landscape scale mosquito control, and render a determination to place it on the appropriate animal import list. Requires DOA, DOH, and DLNR to collaborate on a report to the Legislature with recommendations for appropriate vector control programs.”
(https://www.capitol.hawaii.gov/session2019/bills/HB297_SD1_.htm)

Additionally, House Resolution (HR) 95 passed the Hawaii State House in 2021 urging DLNR, DOA, DOH and UH to implement a mosquito control program using *Wolbachia* to reduce mosquito population levels throughout the state.
(https://www.capitol.hawaii.gov/session2021/bills/HR95_HD1_.htm)

It should be noted that this project has been developed with the full support of, and will be implemented in close coordination with, the Hawaii Department of Health Vector Control Branch. Per Hawaii Revised Statutes §26-13, the Department of Health “shall administer programs designed to protect, preserve, care for, and improve the physical and mental health of the people of the State.” DOH has the authority in Hawaii relating

to mosquitoes and public health, and their staff have decades of expertise to implement mosquito surveillance, control and abatement programs.

Proposed Required Conditions for Importation via HDOA List of Restricted Animals Part A

Included are proposed conditions, suggested in collaboration with the HDOH Vector Control Branch, that could be required for importation if *Culex quinquefasciatus* mosquitoes are added to the HDOA List of Restricted Animals Part A to ensure any future imports meet safeguards to preserve public health, the environment, and the long-term efficiency of the IIT tool. All of the following suggested requirements would need to be met to obtain importation permitting.

Culex quinquefasciatus

1. Only mosquitoes originating from a Hawaii stock are allowed for importation.
2. Only mosquitoes containing the same wild-type bacteria as is already present in Hawaii, or a sexually incompatible *Wolbachia* bacteria compared against Hawaii’s wild mosquito populations are allowed for importation.
3. Only adult male mosquitoes are allowed for importation.
4. Only individuals or organizations who have conducted work for EPA registration trials for mosquito biopesticide products and who can provide data on rearing and sorting methodologies are allowed to ship these mosquitoes to Hawaii.

5. Only individuals or organizations listed on the import application are allowed to import/receive these mosquitoes.
6. Only islands with established or incipient wild mosquito populations, as determined by the Hawaii Department of Health's Vector Control Branch, are allowed to import these mosquitoes.
7. All environmental review processes, including potential Environmental Impact Statements, Environmental Assessments, or other environmental compliance requirements as outlined by State Law and OEQC, must be completed or cited prior to importation.

PQB NOTES: PQB has taken DLNR's proposed permit conditions and incorporated them into the conditions in section VI.

Specific details for importation

This is an application for:

- A permit to import male, mosquito species: *Culex quinquefasciatus*.
- The listing of these mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

Within *Culex quinquefasciatus*, the strain of incompatible bacterium will be *Wolbachia wAlbA*, *Wolbachia wAlbB*, or *Wolbachia wPip4*. These *Wolbachia* bacterium are not present within the corresponding species of Hawaii's established mosquito population. The presence of this bacterium will make these males sexually incompatible with the wild, established female mosquitoes. Once imported, the male, sexually incompatible males will be released according to EPA and HDOA label directions to suppress the population of the established mosquito populations. Based on the prior use of this technology in California, Florida, and Kentucky, there are no data to suggest releases of these male mosquitoes to have a negative impact on agriculture, the environment, or public health and safety. Existing wild-type bacteria strain that may be imported is wPipV, which is already found on all of the main Hawaiian islands.

DISCUSSION:

1. Persons Responsible:

DLNR Chairperson, Suzanne Case
DOFAW Administrator, David Smith
DOFAW Entomologist, Cynthia King
Department of Land and Natural Resources – Oahu
1151 Punchbowl Street, Honolulu, HI 96813

DLNR-DOFAW, Hawaii Invertebrate Program Captive Propagation Facility -
Oahu
779 Ulukahiki Street, Kailua, Honolulu, HI 96813, (808) 266-7989

DLNR Waimano Baseyard – Oahu
2680 Waimano Home Road, Pearl City, HI 96782, (808) 266-7989

Kaua'i Branch Manager, Sheri Mann, Division of Forestry & Wildlife, 3060 Eiwa
Street Rm. 306, Lihue, HI 96766. (808) 274-3433

O'ahu Branch, Division of Forestry & Wildlife, 2135 Makiki Heights Drive,
Honolulu, HI 96822. (808) 973-9778

Maui (& Moloka'i) Branch, Division of Forestry & Wildlife, 1955 Main Street,
Room 301, Wailuku, HI 96793. (808) 984-8100

Hawai'i Branch, Division of Forestry & Wildlife, 19 E. Kawili Street, Hilo, HI
96720. (808) 974-4221

2. Locations and Safeguards:

All mosquitoes for import will originate from Hawaii biotypes collected from

Hawaii. All mosquitoes will be backcrossed for at least 7 generations to ensure >99% Hawaii genetics are contained within the commercially available products to be applied within Hawaii. This backcrossing will also mitigate the risks of infections microorganisms and parasites to the mosquitoes via vertical transmission – thus lowering the risk of the mosquitoes accidentally introducing a new parasite or pathogen. In order for these mosquitoes to acquire and vector a disease, an adult female must blood feed from a disease infected vertebrate, and the pathogen must survive in the mosquito and be injected into another vertebrate during a subsequent blood feeding. As the intended importation of these mosquitoes only includes the importation of male mosquitoes that do not bite or feed on blood, the unintended importation of an acquired pathogen is eliminated. Verification of Hawaii biotypes and *Wolbachia* strains will be conducted on initial shipments of male mosquitoes to verify requirements have been met, in collaboration with University of Hawaii and Department of Health.

These mosquitoes will be imported into Hawaii through the use of commercial cargo flights. Upon reception to Hawaii, the male mosquitoes will be directly released into the laboratory for quality control testing, and into the environment for the purpose of suppressing the wild mosquito populations. These releases will be performed by individuals or organizations certified to apply these mosquito pesticide products to ensures that the product will be applied properly according to the recommended guidelines.

MosquitoMate and Verily will regularly sample release containers by releasing the contents into lab cages and then examining mosquito sex and number. There is an EPA reviewed value of 1 female release per 250,000 males with the MosquitoMate product. A similar value is likely to be estimated for *Culex quinquefasciatus* given that similar automation, engineering and machine learning technology is being applied to sex sorting. MosquitoMate and Verily have not previously identified a female in a single release container during the course of the Puerto Rico or Fresno projects. In another example, a published study estimates the probability at less than 1 female per 200 million males (Crawford JE, Clarke DW, Criswell V, Desnoyer M, Cornel D, Deegan B, et al. Efficient production of male *Wolbachia*-infected *Aedes aegypti* mosquitoes enables large-scale suppression of wild populations. Nat Biotechnol. 2020;38(4):482-92.) To date, PCR monitoring of mosquitoes collected from release field sites have not identified any ZAP infected females.

At least once per year, MosquitoMate and Verily will also conduct longevity and competitiveness studies, comparing the mosquitoes proposed for releases and wild type males. Data from previous trials demonstrate ZAP mosquito longevity and competitiveness to be at least equal to Wild Type males. In addition to Hawaii's import requirements, the shipper and/or receiver will obtain additional permits as required by federal or state agencies.

Wolbachia is an obligate endosymbiont and cannot survive outside of the host invertebrate. *Wolbachia* strains already exist in Hawaii in a range of invertebrates in the wild, including mosquitoes. The presence of *Wolbachia* endosymbionts is the normal state for 40% to 60% of Arthropods and does not represent an unusual or pathogenic bacterial infection. *Wolbachia* are not capable of infecting human cells. MosquitoMate and Verily will perform PCR testing on the mosquitoes to confirm the presence of the correct *Wolbachia* bacterium within the shipment lineage to ensure cytoplasmic incompatibility.

The likelihood that introduced strains of *Wolbachia* would become the dominant strains in the environment is highly unlikely. Replacing the dominant *Wolbachia* strain has been done purposefully in the environment for projects that are separate from the approach we are proposing (such as by the World Mosquito Program in Australia and other nations). To clarify, DLNR is NOT proposing a World Mosquito Program type project where the goal is to intentionally force a different dominant *Wolbachia* strain into the wild mosquitoes in the environment and change vector competence of the wild population. However, in these types of programs, they have to release 4 million mixed male AND female mosquitoes in a given location to force a new *Wolbachia* strain to become the dominate strain over an area of 66 km². Given the aforementioned EPA reviewed value of 1 female release per 250,000 males with the MosquitoMate product, such an outcome is not expected to occur.

If, somehow population replacement were to occur (despite the estimated 1 female release per 250,000 males) DLNR would cease releases as the released males would then be able to mate with the wild females with the established *Wolbachia* species. The outcome of this would be that the mosquito species that already exists in Hawaii would continue to exist in the wild, just with a different *Wolbachia* bacteria. We do not anticipate a different *Wolbachia* bacteria having any new or negative effects on the environment.

DLNR and DOH feel comfortable utilizing these mosquitoes at a very small scale (in remote forest habitat) or at a very large scale (across urban areas and island wide) so long as recommended application guidelines are followed. The scale and scope of the project will likely vary across time based on the funding available and mosquito prevalence. As with any pesticide product, if you do not eradicate the species of concern, they will rebound if you stop using the pesticide product. However, we view this as a beneficial aspect of the project as we also know we can stop the process at any time. Unfortunately, due to the critical nature of the declines of Hawaiian forest birds, we anticipate mosquito control becoming a long-term management action to be performed (similar to rat control and invasive weed control) annually.

Data collection will occur during releases using the State general funds as well as federal funds from partner agencies (USFWS, USGS, NPS), depending on who is performing the releases. As the application of the pesticide product is intended for the reduction of *Culex quinquefasciatus* mosquito populations, this monitoring will include extensive mosquito population surveillance following releases to ensure that populations are reduced. DLNR is already conducting this type of monitoring in preparation for incompatible mosquito releases. *Wolbachia* genetic monitoring will also occur, likely in partnership with USGS, throughout the release program.

In addition to Hawaii's import requirements, the shipper and/or receiver will obtain additional permits as required by federal or state agencies.

3. **Method of Disposition**

Any dead imported mosquitoes will be disposed of as municipal waste.

4. **Abstract of Organism**

Culicidae species are sexually reproducing species. Minimum generation times vary but are approximately three weeks. Mature adults are up to approximately a centimeter in length and can live for a month to a few months. Adult mosquitoes

range from 2.0 to 10.0 mm in size with males being smaller than females on average. Mosquito life cycles are well understood for most species, including all those established in Hawaii.

Larvae feed on organic material found in pools of water. Both adult males and females feed on water that contains carbohydrates (water with sap or nectar). Only mature females of certain species seek out and feed on vertebrate blood prior to egg laying. This blood feeding process allows for the transmission of pathogens and parasites.

Culex quinquefasciatus rely on pools of water with organic material for the growth of larvae. Only adult females bite, as they require blood meals from vertebrate hosts to develop their eggs.

5. Potential Impact to the Environment

Culex quinquefasciatus are already well established in the wild on all of the main islands in Hawaii from sea-level to ~6,000 feet in elevation. and *Culex quinquefasciatus* are established statewide and is well establish on Hawaii's Big Island. An additional five other "biting" non-native mosquito species have also become established: *Ae. albopictus*, *Ae. aegypti*, *Ae. japonicus*, *Ae. vexans*, and *Wyeomyia mitchelli*.

Wolbachia are not infectious to humans and are vertically transmitted through the eggs from one generation to another. The *Wolbachia* bacteria are obligate endosymbionts and can only survive inside the insect host's cytoplasm. A mosquito transinfected with a different strain of *Wolbachia* that results in cytoplasmic incompatibility would not be able to successfully reproduce with a wild mosquito due to cytoplasmic incompatibility. Therefore, if individual mosquitoes did become temporarily established, then they will quickly die off over the following generations because of cytoplasmic incompatibility with wild mosquitoes of the same species, with which they would be expected to encounter and mate.

Through the importation we intend to only import male mosquitoes. The sex separation can be performed in a variety of manners including through computer recognition and separation of males and females or through pupal sorting of males and females. However, if both sexes of transinfected mosquito were to be accidentally released, they are unlikely to maintain a breeding population of a transinfected mosquito. *Wolbachia* invasions into populations require a critical threshold frequency of infection that needs to be overcome before a novel *Wolbachia* infection can spread into a population. The *Wolbachia* infection rate must exceed 20-45% before it can spread and become established. This is evident in large scale releases such as in Cairns, Australia, where millions of transinfected mosquitoes (both sexes) with *Wolbachia* are released into the

environment to control disease transmission, yet they do not easily reach fixation in the wild. If transinfected mosquitoes were to become established, the establishment is likely to be spatially localized due to incompatibility with neighboring mosquito populations.

6. Potential Impacts of Importation

pro: Importation of male mosquitoes will allow the implementation of an evaluated technology that has been scientifically demonstrated as a safe and effective control method for mosquitoes on a landscape-scale. These are mosquitoes that are widespread in Hawaii and which have negative impacts to humans, wildlife, and pets, and are causing the extinction of native forest birds. Thirty species of main Hawaiian forest birds have become extinct since European contact, and another 11 of the 21 remaining species are federally listed as threatened or endangered. The remaining 21 forest bird species remain at great risk as a result of avian pox and avian malaria. Four honeycreeper species (*Akikiki*, *Oreomystis bairdi*; Akekee, *Loxops caeruleirostris*; Kiwikiu, *Pseudonestor xanthophrys* and Akohekohe, *Palmeria dolei*) are of particular concern – each are federally endangered, single-island endemics with highly restricted ranges, number fewer than 1,800 individuals, and display recent alarming population declines. DLNR and USFWS have previously attempted to address these declines through bold conservation actions, such as translocations

and establishment of captive populations; however agencies have met with only limited success due to rapidly changing disease-transmission conditions on the landscape. There is an urgent need to develop new conservation tools, including landscape-level mosquito control in order to prevent further extinctions.

The application of traditional chemical controls for mosquitoes in both natural areas is impractical and causes unacceptable non-target impacts, whereas IIT carries no non-target risks to native species, humans or the environment. Furthermore, mosquitoes were first introduced to the Hawaiian Islands in the 1800s, and while they are used opportunistically as prey items, no species native to Hawai'i are dependent on their presence for survival. The control of mosquito populations in Hawaiian forests would thus cause no negative impacts on Hawaiian species.

Demonstrated application of this approach in Hawaii would have also have a wide range of potential positive effects in that it may facilitate the incompatible insect technique approach being used for human health.

con: It is hard to imagine any negative effects since the species is already established in Hawaii. Importing these organisms will not have any foreseeable beneficial effect to this mosquito species already found in Hawaii. The introduction of, for example, increased genetic variation within the mosquito

species will be minimized by crossing the lines to mosquitoes originating from Hawaii.

The presence of unintended accompanying microbiota is minimized by the sterile laboratory rearing conditions used. These mosquitoes have been maintained for many generations in the lab environment and have not had the opportunity to obtain pathogens from the wild from blood feeding. The presence of intended microbiota, the *Wolbachia*, potentially has very positive effects on the societal health, the suppression of human disease vectored by mosquitoes, the environment, via population suppression of mosquitoes that vector avian pathogens, and the economy, through the potential increased tourism and lessened disease burden.

This mosquito species is already well established in Hawaii, as are many different strains of *Wolbachia*. MosquitoMate and Verily have a demonstrated track record of success utilizing sex-sorting methods which are highly effective. In the event that technical difficulties did occur during sex-sorting methods, because of cytoplasmic incompatibility, the escape of female mosquitoes carrying a new *Wolbachia* strain is not expected to be stable over the following generations. Laboratory reared females outcrossing to locally established wild male mosquitoes will result in cytoplasmic incompatibility and the failure of offspring to develop.

There is an extensive body of literature surrounding this mosquito species, its impact upon Hawaii, and *Wolbachia*-mediated cytoplasmic incompatibility.

References:

Information on *Wolbachia*, with a focus on cytoplasmic incompatibility within mosquitoes:

- Atyame, C. M., Cattell, J., Lebon, C., Flores, O., Dehecq, J.-S., Weill, M., Gouagna, L. C. & Tortosa, P. (2015) *Wolbachia*-based population control strategy targeting *Culex quinquefasciatus* mosquitoes proves efficient under semi-field conditions. *PLoS ONE* 10, e0119288.
- Atyame, C. M., Labbé, P., Lebon, C., Weill, M., Moretti, R., Marini, F., Gouagna, L.C., Calvitti, M. & Tortosa, P. (2016) Comparison of irradiation and *Wolbachia* based approaches for sterile-male strategies targeting *Aedes albopictus*. *PLoS ONE* 11, e0146834.
- Barton, N. H., & Turelli, M. (2011). Spatial waves of advance with bistable dynamics: cytoplasmic and genetic analogues of Allee effects. *The American Naturalist*, 178(3), E48-E75.
- Blagrove, M. S., Arias-Goeta, C., Failloux, A. B., & Sinkins, S. P. (2012). *Wolbachia* strain wMel induces cytoplasmic incompatibility and blocks

- dengue transmission in *Aedes albopictus*. *Proceedings of the National Academy of Sciences*, 109(1), 255-260.
- Callaini, G., Dallai, R., & Riparbelli, M. G. (1997). Wolbachia-induced delay of paternal chromatin condensation does not prevent maternal chromosomes from entering anaphase in incompatible crosses of *Drosophila simulans*. *Journal of Cell Science*, 110(2), 271-280.
- Dobson, S. L., Marsland, E. J., & Rattanadechakul, W. (2001). Wolbachia-induced cytoplasmic incompatibility in single- and superinfected *Aedes albopictus* (Diptera: Culicidae). *Journal of Medical Entomology*, 38(3), 382-387.
- Hamm, C. A., Begun, D. J., Vo, A., Smith, C. C., Saelao, P., Shaver, A. O., ... & Turelli, M. (2014). Wolbachia do not live by reproductive manipulation alone: infection polymorphism in *Drosophila suzukii* and *D. subpulchrella*. *Molecular Ecology*, 23(19), 4871-4885.
- Hoffmann, A. A., Montgomery, B. L., Popovici, J., Iturbe-Ormaetxe, I., Johnson, P. H., Muzzi, F., ... & Cook, H. (2011). Successful establishment of Wolbachia in *Aedes* populations to suppress dengue transmission. *Nature*, 476(7361), 454.
- Hoffmann, A. A., Ross, P. A., and Rašić, G. (2015) Wolbachia strains for disease control: ecological and evolutionary considerations. *Evolutionary Applications*, 8(8), 751-768.
- Jiggins FM. (2017) The spread of Wolbachia through mosquito populations. *PLoS Biology* 15(6):e2002780.
- Laven, H. (1967). Eradication of *Culex pipiens fatigans* through cytoplasmic incompatibility. *Nature*, 216(5113), 383-384.
- Liao W, Atkinson CT, LaPointe DA, Samuel MD (2017) Mitigating Future Avian Malaria Threats to Hawaiian Forest Birds from Climate Change. *PLoS ONE* 12(1):e0168880.
- Mains, J. W., Brelsfoard, C. L., Rose, R. I. & Dobson, S. L. (2016) Female adult *Aedes albopictus* suppression by Wolbachia-infected male mosquitoes. *Scientific Reports* 6, 33846.
- Stouthamer, R., Breeuwer, J. A., & Hurst, G. D. (1999). Wolbachia pipientis: microbial manipulator of arthropod reproduction. *Annual Reviews in Microbiology*, 53(1), 71-102.
- Sinkins, S. P., Braig, H. R., & O'Neill, S. L. (1995). Wolbachia superinfections and the expression of cytoplasmic incompatibility. *Proceedings of the Royal Society of London B: Biological Sciences*, 261(1362), 325-330.
- Tram, U., & Sullivan, W. (2002). Role of delayed nuclear envelope breakdown and mitosis in Wolbachia-induced cytoplasmic incompatibility. *Science*, 296(5570), 1124-1126.
- Waltz, E. (2016) US reviews plan to infect mosquitoes with bacteria to stop disease. *Nature* 533, 450-451.
- Weinert, L. A., Araujo-Jnr, E. V., Ahmed, M. Z. & Welch, J. J. (2015) The incidence of bacterial endosymbionts in terrestrial arthropods. *Proc. R.*

Soc. B 282, 20150249.

- Werren, J. H., Baldo, L., & Clark, M. E. (2008). Wolbachia: master manipulators of invertebrate biology. *Nature Reviews Microbiology*, 6(10), 741.
- Zug, R., & Hammerstein, P. (2012). Still a host of hosts for Wolbachia: analysis of recent data suggests that 40% of terrestrial arthropod species are infected. *PLoS ONE*, 7(6), e38544.

Wolbachia in Hawaii:

- Atkinson, C. T., W. Watcher-Weatherwax, and D. A. LaPointe. (2016) Genetic diversity of *Wolbachia* endosymbionts in *Culex quinquefasciatus* from Hawaii, Midway Atoll and American Samoa. Technical Report HCSU-074.
- Bennett, G. M., Pantoja, N. A., & O'Grady, P. M. (2012). Diversity and phylogenetic relationships of *Wolbachia* in *Drosophila* and other native Hawaiian insects. *Fly*, 6(4), 273-283.

Other References:

- Anonymous. (2017). To Restore a Mosquito-Free Hawaii. Summary Report of the Workshop to Formulate Strategic Solutions for a "Mosquito-Free Hawaii". Available at: "http://www.cpc-foundation.org/uploads/7/6/2/6/76260637/report_on_mosquito_free_workshop.pdf."
- Anonymous. (2020). Three great years of Debug Fresno. Available at: "<https://blog.debug.com/2020/01/three-great-years-of-debug-fresno.html>"
- Dame, D. A., Curtis, C. F., Benedict, M. Q., Robinson, A. S., & Knols, B. G. (2009). Historical applications of induced sterilisation in field populations of mosquitoes. *Malaria Journal*, 8(2), S2.
- Johnston, D., *et al.* (2016). Notes from the field: outbreak of locally acquired cases of dengue fever—Hawaii, 2015. *MMWR. Morbidity and Mortality Weekly Report*, 65(2); 34–35.
- Kauffman, E., Payne, A., Franke, M. A., Schmid, M. A., Harris, E., & Kramer, L. D. (2017). Rearing of *Culex* spp. and *Aedes* spp. mosquitoes. *Bio-Protocol*, 7(17).
- Mendenhall, I. H., S. A. Tello, L. A. Neira, L. F. Castillo, C. B. Ocampo, and D. M. Wesson. (2012) Host Preference of the Arbovirus Vector *Culex Erraticus* (Diptera: Culicidae) at Sonso Lake, Cauca Valley Department, Colombia. *Journal of Medical Entomology* 49 (5): 1092–1102
- O'Neill, S. L.; Ryan, P. A.; Turley, A. P.; Wilson, G.; Retzki, K.; Iturbe-Ormaetxe, I.; Dong, Y.; Kenny, N.; Paton, C. J. & Ritchie, S. A. (2018) Scaled deployment of Wolbachia to protect the community from *Aedes* transmitted arboviruses. *Gates Open Research* 2: 1-18

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- Richmond JY, McKinney RW, eds. (1999). *Biosafety in Microbiological and Biomedical Laboratories* (4th ed.). ISBN 0-7881-8513-6.
- Scott, T. W. (2005). Containment of arthropod disease vectors. *ILAR Journal*, 46(1), 53-61.
- Takken, W., & Verhulst, N. O. (2013). Host preferences of blood-feeding mosquitoes. *Annual Review of Entomology*, 58, 433-453.
- [USFWS] U.S. Fish and Wildlife Service. (2006) Endangered and Threatened Wildlife and Plants; Determination of Status for 12 Species of Picture-Wing Flies from the Hawaiian Islands. *Federal Register* 71 (89): 26835-26852.
- Winchester, J. & Kapan, D. (2013). History of *Aedes* Mosquitoes in Hawaii. *Journal of the American Mosquito Control Association* 29(2): 154-163.

IV. Environmental Assessment (EA):

Pursuant to a May 2008 Hawai'i Intermediate Court of Appeals decision ([‘Ohana Pale Ke Ao v. Board of Agriculture, 118 Haw. 247 \(Haw. App. 2008\)](#)), the Department of Agriculture's (Department's) import permit process is subject to the requirements of the Hawai'i Environmental Protection Act, chapter 343, Hawai'i Revised Statutes (HRS). Under this decision, the requirement for an EA as a condition of the import permit or related authorization applies in those circumstances where the underlying permit activity for the importation initiates a "program or project" and where the use of state or county funds or state or county lands is involved. When those circumstances are present, as they appear to be when a new organism is used in a new program or project located at a facility located at UHM or UHH (state lands), an EA is required to determine whether the proposed project or program is likely to have a significant impact on the environment. However, certain activities may be eligible for "exemption" under provisions established through the Environmental Advisory Council, provided that the project or program is determined to have little or no impact on the environment.

Analysis of Application re EA: Under the above-cited court decision, the EA requirement is triggered under certain circumstances, including when an applicant proposes an action on state lands that requires agency approval and is not specifically exempted under Chapter 343, HRS. That is the case here. The applicant's request in this instance involves the field-release of *Culex quinquefasciatus* for field release to suppress wild populations of *Culex quinquefasciatus* in the environment. So, agency approval is required for the applicant's proposed action/activity on state lands or sensitive habitats. As PQB understands the court's analysis in the 'Ohana Pale decision, the activity proposed under this permit application would initiate a project that may use state lands and/or sensitive habitats, initially triggering the EA requirement.

DLNR has provided an "Exemption Notice Regarding the preparation of an environmental assessment under the authority of Chapter 343, HRS and Section 11-200.1-17, HAR" and intend to conduct EA's to be completed at future dates.

V. Proposed Permit Conditions:

1. The restricted article(s), Hawaiian biotype Southern House Mosquito, *Culex quinquefasciatus* (Say, 1823), inoculated with a foreign *Wolbachia* bacteria species, shall be used for field-release for area-wide mosquito suppression, a purpose approved by the Board of Agriculture (Board). Live sale or transfer of the restricted article(s) is prohibited, except as approved by the Board. Transport to or release on any island that does not have a population of *Cu. quinquefasciatus* is prohibited.
2. Only male restricted article(s) shall be imported and released.
3. Only Hawaiian biotype *Cu. quinquefasciatus* that have been backcrossed with mosquitos collected in Hawaii for at least 7 generations or 100% Hawaii-collected *Cu. quinquefasciatus* and their progeny, shall be imported.
4. Only restricted article(s) inoculated with *Wolbachia pipientis* bacteria strains already occurring in Hawaii mosquitos or strains wAlbA, wAlbB, wPip4, and wPip5 shall be imported.
5. The permittee, Suzanne Case, Chairperson, State of Hawaii Department of Land and Natural Resources (DLNR), 1151 Punchbowl Street, Honolulu, Hawaii, 96813 shall be responsible and accountable for all restricted article(s) imported from the time of receipt until their final disposition.
6. The restricted article(s) shall be maintained by the responsible DLNR personnel, Suzanne Case, David Smith, or Cynthia King or by trained or certified personnel designated by the permittee. A list of trained or certified personnel shall be provided to the Plant Quarantine Branch (PQB).

PQB NOTES: *Condition 6 amended to reflect the Committee's desire to have a list of trained or certified DLNR personnel.*

7. The restricted article(s) shall be safeguarded at the following sites listed below, inspected and approved by the PQB prior to importation. Movement of the restricted article(s) to another site shall require a site inspection and approval by the PQB Chief prior to movement.
 - a. DLNR Waimano Baseyard – Hawaii Invertebrate Program - Oahu 2680 Waimano Home Road, Pearl City, HI 96782, (808) 266-7989

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- b. Kaua'i Branch, Division of Forestry & Wildlife, 3060 Eiwa Street Rm. 306, Lihue, HI 96766. (808) 274-3433
- c. O'ahu Branch, Division of Forestry & Wildlife, 2135 Makiki Heights Drive, Honolulu, HI 96822. (808) 973-9778
- d. Maui (& Moloka'i) Branch, Division of Forestry & Wildlife, 1955 Main Street, Room 301, Wailuku, HI 96793. (808) 984-8100
- e. Hawai'i Branch, Division of Forestry & Wildlife, 19 E. Kawili Street, Hilo, HI 96720. (808) 974-4221

PQB NOTES: *Condition 7 amended to remove reference to progeny.*

- 8. The restricted article(s) shall be maintained by Suzanne Case, State of Hawaii Department of Land and Natural Resources (DLNR), 1151 Punchbowl Street, Honolulu, Hawaii, 96813, or by trained or certified personnel designated by the permittee.

PQB NOTES: *Condition 8 amended to remove reference to progeny.*

- 9. The permittee shall submit samples of the restricted article(s) prior to importation to the PQB upon request.
- 10. Prior to the arrival of each shipment containing the restricted article(s), the permittee shall provide to the PQB Chief the following information in writing:
 - a. Expected arrival date;
 - b. A copy of the shipping waybill or tracking numbers for each parcel;
 - c. A copy of the invoice, packing list or other similar PQB approved document that states the quantity of the restricted article(s), the scientific and common name(s) of the restricted article(s), the shipper, and the consignee for the restricted article(s);
 - d. The names and addresses of the shipper and permittee; and
 - e. The total number of parcels.
- 11. The restricted article(s) shall be imported only through the port of Honolulu, except as designated by the Board. Entry into Hawaii through another port is prohibited unless designated by the Board.

12. At least four sides of each parcel containing the restricted article(s) shall be clearly labeled in plain view with "Live Animals" and "This Parcel May be Opened and Delayed for Agriculture Inspection", in ½" minimum-sized font.
13. The restricted article(s) shall be shipped in sturdy PQB-approved containers designed to be escape-proof and leak-proof.
14. Each shipment of the restricted article(s) shall be accompanied by a complete copy of the PQB permit with permit conditions for the restricted article(s), and an invoice, packing list or other similar PQB approved document listing the scientific and common names of the restricted article(s), the quantity of the restricted article(s), the shipper, and the permittee for the restricted article(s).
15. The permittee shall immediately notify the PQB Chief in writing under the following circumstances:
 - a. If any escape, theft, accidental release, disease outbreaks, pest emergence and/or mass mortalities involving the restricted article(s), under this permit occurs. The department may confiscate or capture the restricted article(s) and any progeny that escapes or is found to be free from confinement at the expense of the owner, pursuant to the Hawaii Revised Statutes (HRS), §150A-7(c).
 - b. If any changes are made to the approved sites, facilities or containers used to hold the restricted article(s).
 - c. If a shipment of the restricted article(s) is delivered to the permittee without a PQB "Passed" stamp, tag or label affixed to the article, container or delivery order that indicates that the shipment has passed inspection and is allowed entry into the State. Under this circumstance, the permittee shall not open or tamper with the shipment. Additionally, the permittee shall secure all restricted article(s), shipping containers, shipping documents and packing materials for the PQB.
 - d. If the permittee is found in violation of any municipal, state or federal policies, rules and/or laws, pertaining to the restricted article(s).
 - e. If the permittee will no longer import and/or possess the restricted article(s) authorized under this permit. Under this circumstance, the permittee shall inform the PQB Chief of the final disposition for the restricted article(s) and the permit shall be canceled.
16. In the event that the restricted article(s) become parasitized or infected by disease, all restricted article(s) from which the parasitized or infected restricted

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article(s) originated shall be considered compromised and immediately subjected to a treatment(s) approved by the PQB Chief. All shipping containers, packing materials, equipment, and any other items used in conjunction with the compromised restricted article(s), shall also be subjected to a treatment(s) approved by the PQB Chief.

PQB NOTES: *Condition 16 amended to remove reference to progeny.*

17. Prior to interisland transport, all restricted article(s) shall be presented to the PQB for inspection. The permittee shall also follow Permit Conditions Nos. 12, 13, and 14 for each interisland shipment. The PQB inspector shall affix an interisland certificate of inspection to the shipment as verification of a completed inspection.
18. The permittee(s) shall submit an annual report to the PQB on the results of all research including post-release monitoring programs. The report shall be submitted by the 31st of January of each year and shall cover the prior 12-month period. Information reported shall include:
 - a. Number of mosquito releases per site.
 - b. Number of mosquitoes released per site.
 - c. Impact on wild mosquito populations.
 - d. Detections of introduced Wolbachia strains in wild mosquito populations,
 - e. Impact of mosquito releases on native bird populations.
19. The permittee(s) shall adhere to the use, facility, equipment, procedures, and safeguards described in the permit application, and as approved by the Board and the PQB Chief.
20. Any approved site, restricted article(s), and records pertaining to the restricted article(s) under permit may be subject to post-entry inspections by the PQB, upon arrival at the permittee's facility. The permittee shall make the approved site, restricted article(s), and records pertaining to the restricted article(s) available for inspection upon request by a PQB Inspector.
21. The permittee shall have a biosecurity manual available for review and approval by the PQB, at the time of the initial site inspection and any subsequent post-entry inspections, which identifies the practices and procedures to be adhered to by the permittee, to minimize the risk of theft, escape, or accidental release of the restricted article(s), including minimizing the risk of introduction and spread of diseases and pests associated with the restricted article(s) to the environment.

The permittee shall adhere to all practices and procedures as stated in this biosecurity manual.

22. The permittee shall submit to the PQB Chief a copy of all valid licenses, permits, certificates, or other similar documents required by other agencies for the restricted article(s). The permittee shall immediately notify the PQB Chief in writing when any of the required documents are suspended, revoked, or terminated. This permit may be amended, suspended, or canceled by the PQB Chief in writing, upon suspension, revocation, or termination of any required license, permit, certificate or similar document for the restricted article(s).
23. It is the responsibility of the permittee to comply with any and all applicable requirements of municipal, state, or federal law pertaining to the restricted article(s).
24. The permittee shall be responsible for all costs, charges, or expenses incident to the inspection, treatment, or destruction of the restricted article(s) under this permit, as provided in Act 173, Session Laws of Hawaii 2010, Section 13, including, if applicable, charges for overtime wages, fixed charges for personnel services, and meals.
25. Any violation of the permit conditions may result in citation, permit cancellation, and enforcement of any or all of the penalties set forth in HRS §150A-14.
26. A canceled permit is invalid and upon written notification from the PQB Chief, all restricted article(s) listed on the permit shall not be imported. In the event of permit cancellation, any restricted article(s) imported, may be moved, seized, treated, quarantined, destroyed, or sent out of State at the discretion of the PQB Chief. Any expense or loss in connection therewith shall be borne by the permittee.
27. The permit conditions are subject to cancellation or amendment at any time due to changes in statute or administrative rules restricting or disallowing import of the restricted article(s) or due to Board action disallowing a previously permitted use of the restricted article(s).
28. These permit conditions are subject to amendment by the PQB Chief in the following circumstances:
 - a. To require disease screening, quarantine measures, and/or to place restrictions on the intrastate movement of the restricted article(s), as appropriate, based on scientifically validated risks associated with the restricted article(s), as determined by the PQB Chief, to prevent the introduction or spread of diseases and/or pests associated with the restricted article(s).

- b. To conform to more recent Board approved permit conditions for the restricted article(s), as necessary to address scientifically validated risks associated with the restricted article(s).
29. The permittee shall agree in advance to defend and indemnify the State of Hawaii, its officers, agents, and employees for any and all claims against the State of Hawaii, its officers, agents, employees, or Board of Agriculture members that may arise from or be attributable to any of the restricted article(s) that are introduced under this permit. This permit condition shall not apply to a permittee that is a federal or State of Hawaii entity or employee, provided that the State or federal employee is a permittee in the employee's official capacity.

VI. Advisory Review

ADVISORY SUBCOMMITTEE REVIEW: This request was submitted to the Advisory Subcommittee on Entomology for its review and recommendation. Advisory Subcommittee recommendations and comments are as follows:

Provided *Culex quinquefasciatus* is placed on the list of Restricted Animals (Part A), I recommend approval ___/___ disapproval to allow the importation of lab-reared strains of the mosquito, *Culex quinquefasciatus* (Diptera: Culicidae), inoculated with strains of *Wolbachia* bacteria for immediate field release to suppress wild populations of *Culex quinquefasciatus* by the DLNR.

Dr. Daniel Rubinoff: Recommends Approval.

Ms. Janis Matsunaga: Recommends Approval.

Comments: "Provided *Culex quinquefasciatus* is placed on the list of Restricted Animals (Part A), I recommend approval to allow the importation of lab-reared strains of the mosquito, *C. quinquefasciatus* (Diptera: Culicidae), inoculated with strains of *Wolbachia* bacteria for immediate field release to suppress wild populations of *C. quinquefasciatus* by the DLNR given questions I provide in the word document are addressed."

Dr. Mark Wright: Recommends Approval.

Comments: "The applicants have provided documentation showing the stringent quality control systems they have in place to ensure that only male mosquitoes are released. There are data showing that this mosquito suppression method can be highly effective. There is enormous potential benefit of implementing this technology in Hawaii."

Provided *Culex quinquefasciatus* is placed on the list of Restricted Animals (Part A), I recommend approval ___/___ disapproval to establish permit conditions for the importation and immediate field release of lab-reared strains of the mosquito, *Culex quinquefasciatus* (Diptera: Culicidae) inoculated with strains of *Wolbachia* bacteria for immediate field release to suppress wild populations of *Culex quinquefasciatus* by the DLNR.

Dr. Daniel Rubinoff: Recommends Approval.

Comments: "This technology is low risk and should be fast tracked ASAP."

Ms. Janis Matsunaga: Recommends Approval.

Comments: "Provided *Culex quinquefasciatus* is placed on the list of Restricted Animals (Part A), I recommend approval to establish permit conditions for the importation and immediate release of lab-reared strains of the mosquito, *C. quinquefasciatus* (Diptera: Culicidae) inoculated with strains of *Wolbachia* bacteria for immediate field release to suppress wild populations of *C. quinquefasciatus* by the DLNR given the following questions are addressed:

What and where are these EPA and HDOA label directions?

What and where are the recommended guidelines for releasing these mosquitoes?

Will these *Wolbachia* inoculated mosquitoes be classified as a bio-pesticide product in Hawaii?

What quality control testing will be done in the laboratory in Hawaii just prior to release of the imported mosquitoes into the environment?

-How will the results be reported?

-How will the results affect releases into the environment?

Will there be routine monitoring of traps for incompatible adult females or larvae as proposed by Verily Life Sciences?

The applicant is applying for 1 million mosquitoes. How many mosquitoes will be in each shipment? What is the duration of time between shipments?

What is the plan and procedure for releases into the environment?

- How will it be decided how many mosquitoes are leased at each time at each site?
- How will each site of release be determined?
- Is there a potential release site list available?"

PQB NOTES: DLNR's answers to Subcommittee member Matsunaga's questions are listed below. For clarity, Subcommittee member Matsunaga's questions are italicized in blue. DLNR's responses are in black.

What and where are these EPA and HDOA label directions for releasing the mosquitos?

DLNR response: "The EPA is currently reviewing the label for the tool as submitted by HDOA. Below are the DRAFT label criteria:

- i. **Sites to be Treated:** State, Federal and Private wildlife conservation areas that contain *Cx. quinquefasciatus* mosquitoes throughout the State of Hawaii.
- ii. **Method of Application:** Point releases by hand or aerial releases.
- iii. **Rate of Application:** Initial absolute rates of release are pending a Mark Release Recapture in the proposed treatment area as this will establish baseline mosquito populations in the treatment area and ecology, field longevity and other factors used to estimate release rates (number of males/acre/week) sufficient to achieve and maintaining the "overflowing ratio" of $\geq 10:1$ DQB males:wild type (WT) male *Cx. quinquefasciatus* in adult traps in the release area as described on the DQB label. If wild type populations are significantly suppressed release rates may be lowered while still achieving $\geq 10:1$ overflowing DQB:WT males.
- iv. **Maximum Number of Applications:** *156 applications per release site per year based on maximum of 3 releases per week.* At every treatment location release may be for up to a year, with the intention of having multiple releases per week. If strong suppression is achieved releases may be reduced in frequency at a given location with releases starting at a new location at the same cadence. Thus the total number of application days is up to $3 \times 52 = 156$ during the year. If the permit is extended then a similar rate of releases will occur
- v. **Total Acreage to be Treated:** Up to 40,000 acres of State, Federal and Private wildlife conservation areas in the State of Hawaii.
- vi. **Total Amount of Pesticide to be Used:**
Maximum amount of DQB Males to be applied per year: Up to 4,000,000 males per week = 208,000,000 males/year. Maximum amount of *Wolbachia pipientis*, wAlbB to be applied per year: Up to ~25g/week = 1300g/year"

What and where are the recommended guidelines for releasing these mosquitos?

DLNR response: "See above DRAFT label criteria submitted by HDOA."

Will these Wolbachia inoculated mosquitoes be classified as a bio-pesticide product in Hawaii?

DLNR response: "Yes, they will be classified as a bio-pesticide by the EPA and HDOA."

What quality control testing will be done in the laboratory in Hawaii just prior to release of the imported mosquitoes into the environment?

-How will the results be reported?

DLNR response: "QA/QC will be conducted at the rearing facility in California and measurements of the number of dead IIT males will be estimated to inform and perfect the shipping mechanism prior to release. The results of QA/QC will be reported to HDoA/USFWS/DLNR releases. QA/QC of both the Wolbachia type and efficacy should be done outside of Hawaii to reduce any possibility of female releases (and thus lower the tools efficacy)."

-How will the results affect releases into the environment?

DLNR response: "If QA/QC is significantly reduced after shipping then this will lead to a reduced efficacy of the product, which will in turn lead to an increase in projected prevalence of the avian malaria vector *C. quinquefasciatus*. As *C. quinquefasciatus* is an invasive organism reducing its prevalence will not cause undo harm, and being ineffectual will result in no increased impact to native forest birds."

Will there be routine monitoring of traps for incompatible adult females or larvae as proposed by Verily Life Sciences?

DLNR response: "Yes, post release monitoring is essential to maintain the efficacy of the tool. Both female and male presence in traps will be monitored and assessed for Wolbachia type. "

The applicant is applying for 1 million mosquitoes. How many mosquitoes will be in each shipment? What is the duration of time between shipments?

DLNR response: "The applicant (HDoA) is applying for >1 million mosquitoes to be released per week. The number in each shipment as estimated using a 10:1 overflooding ratio and will be determined by the area treated. Shipments could be as often as be bi-weekly depending on wild-type densities upon initial product application."

What is the plan and procedure for releases into the environment?

DLNR response: "Releases into the environment will occur via pedestrian releases, pedestrian assisted aerial releases, and aerial releases. Each of the modes is being included in environmental assessments. Releases will follow aforementioned draft label criteria, as well as standard operating procedures relating to management actions and helicopter operations for DOFAW field staff."

-How will it be decided how many mosquitoes are released at each time at each site?

DLNR response: "This will be determined using a minimum overflooding ratio of 10:1, which has been shown to reduce mosquito prevalence by >90%."

-How will each site of release be determined?

DLNR response: "Sites that are of conservation based priority due to the presence of remaining populations of rare forest birds will be prioritized for release by state and federal agencies to maximize likelihood of species recovery."

-Is there a potential release site list available?

DLNR response: "As stated above the HDoA EPA permit application is for State, Federal and Private wildlife conservation areas that contain *C. quinquefasciatus* mosquitoes throughout the State of Hawaii. Initial releases will be conducted on priority Forest bird conservation sites on Maui, followed by Kauai and then the Big Island of Hawaii. Proposed project areas are to be specifically identified in environmental assessment documents. Initial public scoping for the Maui EA has been completed (in advance of publishing the draft EA) and information regarding East Maui project areas can be found here:

[file:///C:/Users/ckingcb/Downloads/HALE_ScopingNewsletter_12.7.21%20\(3\).pdf](file:///C:/Users/ckingcb/Downloads/HALE_ScopingNewsletter_12.7.21%20(3).pdf)

Project areas on Kauai, Big Island have not yet been finalized. However, a statewide EA will be completed which will cover environmental compliance for all state and private lands across Hawaii."

Dr. Mark Wright: Recommends Approval.

Comments: "This application makes a strong case for these releases to be made. Environmental risks of taking this action are minimal."

ADVISORY COMMITTEE REVIEW:

This request was reviewed by the Committee on June 9, 2022. Based on the large number of written testimonies that were submitted Chairperson Darcy Oishi took oral testimony prior to hearing the submittals to ensure that we have received written testimony for both mosquito proposals being review by the committee today AND the Board order. He said because it is difficult to separate the two based upon how the written testimony was collected, all testimony would be considered together.

Mr. Chris Farmer, Hawaii Program Director for the American Bird Conservancy spoke in strong support for the recommendation to list the three mosquito species. He said

Hawaii is experiencing a conservation crisis and the *Wolbachia* method that DLNR and DOH have proposed is the best, safest solution to save these birds. He noted all three mosquitoes are found widely in the State and that the bacteria are here and is widely spread as well. He expressed concerned hearing there is opposition claiming the mosquitos are genetically modified because this is not a genetic modification technique. There is no GM modification. There is no manipulation of any of the genome. Mr. Farmer said the technique is reversable, so there is the ability to go very safely and slowly if needed. He said the technique is widely used, and safely used for human health around the globe and on the continent. He said this is the best option to save the Hawaiian honey creepers and our native birds. He urged a recommendation to approve the DOH and DLNR requests.

Ms. Christy Martin, Coordinating Group on Alien Pest Species, strongly supports both proposals. She reiterated Mr. Farmers' sentiment saying native birds are running out of time. She said this is the only available technique to try to save some of these birds. She noted the non-native zebra doves are the ones really infected with diseases such as avian malaria and avian pox and when the mosquitoes go and bite them, they can transfer the diseases to the native birds. She urged the Committee to recommend both submittals be approved.

Ms. Chelsea Arnott, Hawaii Invasive Species Council said they submitted written testimony in support of both submissions. She noted the importance of having large-scale tools developed and deployed to help prevent the spread of invasive species. She said climate change is exacerbating the extinction of native forest birds. She said this tool and technique is being deployed internationally and nationally to control and reduce large population of mosquitoes not only for human health but for natural resources. She said this is the best and safest tool for reducing mosquitoes, saving forest bird from extinction, and protecting human health.

Ms. Ulalia Woodside, Executive Director of the Nature Conservancy, said the Nature Conservancy strongly supports the DLNR and DOH applications regarding the mosquito species. She said these permits will allow Hawaii to take steps towards developing and implementing a mosquito control program that uses *Wolbachia* bacteria to reduce mosquito populations throughout the state which would have a positive impact on human health and the precarious state of native forest birds. She said the work with *Wolbachia* is not a genetically modified organism or any type of genetic engineering. She noted the expansion of mosquitos is causing arapid decline in native forest bird populations and if action isn't taken, there will be many more extinctions noting it may already be too late for some. She said this is the way to save Hawaii's native forest birds.

After the oral testimony, Mr. Christopher Kishimoto, PQB Entomologist, provided a summary of the request. During the summary, Mr. Kishimoto noted that Subcommittee member Janis Matsunaga brought up a number of questions in one of her responses.

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He noted that DLNR's responses were not received before packet distribution to the Committee, so read DLNR's responses to each of Ms. Matsunaga's questions.

PQB NOTES: *DLNR's exact responses to Ms. Matsunaga's questions can be found on pages 26-28.*

Committee member Dr. Samuel Gon referred to a comment earlier in the testimony that this was a tool used for control of mosquitos in various locations internationally as well as in the United States. He noted that the DLNR proposal may be the very first proposal to use *Wolbachia* for the purpose of conserving natural resources as opposed to a human health purpose. He said it would be an exciting precedent for the use of this tool as a natural resource management tool.

Chairperson Darcy Oishi said Ms. Matsunaga's comments said got him thinking about transportation issues, noting permit condition 11 requires importation through the port of Honolulu and asked if DLNR had comments on any issues they might anticipate. He surmised transportation from U.S. mainland production sites to Hawaii are going to be significantly longer. He noted releases are on Maui and if everything has to go through the port of Honolulu, would this impact the efficacy of the program? He asked DLNR if this permit condition is too restrictive? He asked PQB if the intent is only for males to be introduced and released, is there a need to route everything through the port of Honolulu?

Mr. Kishimoto said usually, most imports of live animals already go through Honolulu as the first port of entry because it is where PQB has the most control over imports. He said Maui and Kauai ports are not considered full ports of entry and they can't receive things like this for inspection. Chairperson Oishi noted that a Board order could change that. Mr. Kishimoto said he wasn't sure if a Board order could change that, but said it definitely is a decision made by the Board. Chairperson Oishi asked how will these insects be moved? He assumed air freight, but could it be a parcel carrier like Fed Ex and UPS? Mr. Kishimoto said that is probably something that DLNR can best answer.

Committee member Cynthia King said she actual can't answer the question about the actual carriers because it has not been discussed. Referring to Mr. Oishi's comment about the additional time where mosquitos have to transit through the port of Honolulu to undergo inspection then move on to the neighbor islands does increase the time they are in less ideal conditions, putting them at a greater risk of higher mortality and lower survivorship or effectiveness in the field. She followed up that it's not to say that DLNR can't do it, but if there was a means by where the port of entry could be altered, DLNR would certainly be supportive of that.

Chairperson Oishi asked what the farthest distance Verily or MosquitoMate has moved mosquitos by air for release? He clarified he wasn't necessarily referring only to distance, but also the time from collection from the rearing facility to release to the field. Ms. King said the longest distance she is aware of is from California to Puerto Rico. She said the total time the mosquitos can be in transit is about 24 hours give or take, but less time in tiny, chilled containers is typically better.

Chairperson Oishi asked if adding on movement from Honolulu to any other island could potentially push transportation times beyond the 24-hour. Committee member Joshua Fisher answered that remains to be determined. He said for the Puerto Rico applications, Fed Ex worked. He said you want to do shipments within 24 hours, and there may be an upper limit to that. He said various delivery services will have to be looked at to best minimize the travel distance. He noted mosquitos coming from California will not be in transit as long versus shipments coming from the east coast, but that would need to be determined and all options are on the table.

Jonathan Ho, PQB Inspection and Compliance Section Chief, said the Board could determine another port of entry but that creates a logistical problem with regards to inspection, particularly because PQB does not have a lot of staff on the neighbor islands. He said the mode of entry plays a specific role. For example, FedEx only comes through the port of Honolulu and there is no direct flight to an outer island and if you are doing air cargo that would be different. He said that PQB deals with live animal shipments all the time and are all required to come through Honolulu, and they are all done well within 24 hours particularly now that many of the COVID restrictions are being loosened. He said an example could be a shipment that comes in through Hawaiian Cargo for Kauai would be presented for inspection on Oahu. As soon as the inspection is completed, the cargo is on the next plane going to that island. He said the duration would be between one and three hours depending on the flight time but completing movement within 24 hours is all but guaranteed to occur provided that there is not some crazy shipping issue. He noted the port of entry issue is somewhat complicated.

Committee member Dr. Maria Haws noted as a minor point that there are two misspellings in the first two clauses related to the subject request and it looks like the second usage of the species name has been misspelled.

Committee member King noted the 24-hour time period isn't limited only to getting a parcel physically to the neighbor island but needs to include the time to get out into the field and then deployed, and that might entail hiking out on foot in some areas where the helicopter or other aeriels deployment that remains to be seen that could be faster in some cases. She wanted to point out there is potentially additional transit time to get the mosquitos to remote field types where the last of our remaining forest birds take refuge.

Chairperson Oishi said there may some potential conflicts in the permit conditions. If only males are imported, references to “progeny” should be removed from permit conditions 8 and 16. Mr. Kishimoto said that can be done. Chairperson Oishi referenced permit condition 6 where it says maintenance by “trained or certified personnel designated by the permittee.” He asked if it’s PQB’s intent to know who is certified and trained and if so, should the permit condition language be modified to reflect reporting, or is it going to be captured within the bio-security plan requirement? Mr. Kishimoto said it is nice to know who the trained staff is, and they can supply PQB with a list, but it doesn’t necessarily have to be a list, it can be inside their reports as well. Chairperson Oishi said it would be cleaner to modify the permit condition language so the applicant knows the expectation. He asked if that would be a problem with DLNR? Chairperson Oishi said he saw Ms. King shake her head indicating “no”.

Chairperson Oishi noted the submittal indicated DLNR will be doing an EA at a future date and heard during Mr. Kishimoto’s presentation that partial EA was done for Maui. He asked what the timeline for completion of the EA was. Committee member King said the timeline for the draft EA to be released to the public is in September of this year for the East Maui area. She said there is a parallel process going on for the proposed Hawaii project area and then there will be a third EA to cover statewide, so a total of three EA’s completed for this.

Chairperson Oishi said the abundance of written testimony against this proposal is tied to concerns over GMO or genetically engineering and assumed the EA will help address some of those concerns. Ms. King said the goal is to put it through the process and make it publicly available so that folks have the opportunity to learn about the project and express any concerns that they might have. I’m sorry I just checked my She corrected her previous statement noting the draft EA is actually supposed to be published in August, not September.

Chairperson Oishi thanked Ms. King for the clarification and asked if there be other outreach or education opportunities to get the word out that this is not using genetically modified organisms. He asked if DLNR has a plan to educate the general public about what this really is. Ms. King said DLNR is part of a multi-agency partnership called Birds Not Mosquitos and while DLNR doesn’t specifically have outreach staff directed to this project at this time we are a partner in this consortium which has emphasized a great deal of outreach and education to the public as well as legislators and community members and it has been an emphasis for a long time. She said there is still a lot of work to do out there based on some of the testimonies that we saw today from concerned members of the public, but noted it was understandable. She said recently there was additional funding allocated towards implementing incompatible insect

techniques in Hawaii and a portion of those funds will certainly be going to continuing outreach.

Committee member Gon commented that DLNR's Dan Dennison released a news release that indicated that there were no GMOs or GEs involved in this particular proposal a couple of days ago and thought it's part of the ongoing multi-pronged outreach and public awareness efforts that going on in the Bird Not Mosquitos group. Ms. King thanked Dr. Gon and noted DLNR has issued other press releases at different stages as the interest in *Wolbachia* for forestry conservation has evolved.

Committee member Joshua Fisher said after reading throughout all the testimonies that were received, he was going to recommend to the communication group working on outreach that they are going to have to broaden to different audiences because it seemed there was a very large farming contingent, or farming community, that had submitted testimony. He felt that outreach could be extended to those audiences. Dr. Gon commented that it is interesting that the rise of anti-GMO sentiment in Hawaii was largely out of the farming and pesticides community, therefore, maybe it isn't that surprising.

Chairperson Oishi asked the Committee if there was any other points of discussion? Hearing none, he asked for a motion. Committee member Sam made a motion to approve this request with the corrections and modifications that Chair Oishi and Dr. Haws mentioned. Chairperson Oishi referenced the changes to permit condition 8 and 16 to eliminate the word "progeny" and permit condition 6 to specify the applicant is to provide a list of individuals that have received training, and the typographic errors in the title. Dr. Gon agreed. Committee member Haws seconded the motion.

Chairperson Oishi asked if there was any further discussion. Committee member King noted that the permit conditions reference quality control upon arrival at University of Hawaii. She believed that was a condition from the previous permit but didn't believe it was included in this request because of the direct release approach. She wanted to verify if it's on the application. She didn't have any concerns about it, but if it was a carryover, she would like to see it corrected.

After discussion it was determined that quality assurance at the University of Hawaii was not included, and Ms. King confirmed that it should not be part of the permit conditions because the University of Hawaii is not aware of any quality assurance requirements. Chairperson Oishi noted permit condition 7 also refers to progeny. Mr. Kishimoto understood.

Chairperson Oishi asked if there was any further discussion. Hearing none, he called for the vote.

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Culex quinquefasciatus
Field Release
Suzanne Case, DLNR


Board of Agriculture

Vote: Approved 7/0.

Motion carries.


STAFF RECOMMENDATION: Based on the favorable responses by the responding Advisory Subcommittee Members' responses and the Advisory Committee's unanimous (7-0) recommendation to approve this request, the Plant Quarantine Branch recommends approval of this request provided the Board has approved the placement of *Culex quinquefasciatus* on the List of Restricted Animals, Part A.

Respectfully Submitted,



BECKY AZAMA
Acting Manager, Plant Quarantine Branch

CONCURRED:



HELMUTH ROGG, Ph.D.
Administrator, Plant Industry Division

APPROVED FOR SUBMISSION:



PHYLLIS SHIMABUKURO-GEISER
Chairperson, Board of Agriculture

PQ-7 (01/04)



State of Hawaii
Department of Agriculture
PLANT QUARANTINE BRANCH
1849 Auiki Street, Honolulu, HI 96819 3100
Phone: (808) 832 0566, FAX: (808) 832 0584

**PERMIT APPLICATION FOR
RESTRICTED COMMODITIES
INTO HAWAII**

<i>For Office Use Only</i>	
Fee: \$ _____	Receipt No. _____
<input type="checkbox"/> Approve Permit No. _____	Date: _____
<input type="checkbox"/> Disapprove	<input type="checkbox"/> Other _____
Processed by: _____	Date: _____

Date: 4/1/22

In accordance with the provision of Chapter 4 71, Hawaii Administrative Rules of the Division of Plant Industry, Department of Agriculture, a permit is requested for the following commodities:

Please type or print clearly.

Quantity	Commodity	Scientific Name
1,000,000	Southern House Mosquito	Culex quinquefasciatus

Name and address of shipper: Department of Land and Natural Resources, Division of Forestry and Wildlife
1151 Punchbowl Street, Honolulu, HI 96813

(Mainland or Foreign address)

Approximate date of arrival: TBD

Please type or print clearly.

Mode of Shipment: Mail Air Freight Boat

Applicant's Name Suzanne Case

Type of Permit:

--- Import
 one time only multi-shipments

--- Intrastate shipment
 one time only multi-shipments

Possession

Company Name Department of Land and Natural Resources
(if applicable)

Hawaii Mailing Address 1151 Punchbowl Street, Honolulu, HI 96813

Telephone number 808 587 0400

Object of importation:

Kept caged at all time
 Used for propagation
 Imported for exhibition
 Imported for liberation
 Other purposes - specify _____

Facsimile number _____

Fee Amount Enclosed (cash, check or mail order) \$ Journal Vouch

(complete reverse side)

Subcommittee on Entomology

G. Simmons & L. Wells – Hawaii Department of Health

D. Smith & C. King – Hawaii Department of Land and Natural Resources

<https://hdoa.hawaii.gov/wp-content/uploads/2019/08/Plant-and-Non-Domestic-Animal-Quarantine-Non-Domestic-Animal-Import-Rules.pdf>

Date:4/1/2022

To:

Advisory Subcommittee on Entomology

From:

Suzanne Case

Department of Land and Natural Resources

1151 Punchbowl Street, Honolulu, HI 96813

David G. Smith

Hawaii Department of Land and Natural Resources, Division of Forestry and Wildlife

1151 Punchbowl Street, Room 325, Honolulu, HI 96813

Cynthia King

Hawaii Department of Land and Natural Resources, Division of Forestry and Wildlife

1151 Punchbowl Street, Room 325, Honolulu, HI 96813

Summary Description of the Requests

In accordance with the provisions of Chapter 150A, Hawaii Revised Statutes, we are requesting to import the following animal commodities:

Commodity	Scientific Name	Quantity
Southern House Mosquitoes (Male Adults)	<i>Culex quinquefasciatus</i>	Continued shipments for immediate release.

Additionally, we are requesting the listing of *Culex quinquefasciatus* mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

Reason for importation:

For immediate field release applications to suppress mosquito populations in areas where Hawaii fauna are at risk of disease transmission due to the presence of these mosquitoes.

Shippers:

- 1) Stephen Dobson, MosquitoMate, Inc.
2520 Regency Rd., Lexington, KY, 40503
- 2) Verily Life Sciences
269 E Grand Ave, South San Francisco, CA 94080

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Importers:

- 1) DLNR Waimano Baseyard – Hawaii Invertebrate Program - Oahu
2680 Waimano Home Road, Pearl City, HI 96782, (808) 266-7989
- 2) Kaua'i Branch, Division of Forestry & Wildlife, 3060 Eiwa Street Rm. 306, Lihue, HI 96766. (808) 274-3433
- 3) O'ahu Branch, Division of Forestry & Wildlife, 2135 Makiki Heights Drive, Honolulu, HI 96822. (808) 973-9778
- 4) Maui (& Moloka'i) Branch, Division of Forestry & Wildlife, 1955 Main Street, Room 301, Wailuku, HI 96793. (808) 984-8100
- 5) Hawai'i Branch, Division of Forestry & Wildlife, 19 E. Kawili Street, Hilo, HI 96720. (808) 974-4221

Project:

This is an application for:

- A permit to import male *Culex quinquefasciatus* mosquito species.
- The listing of *Culex quinquefasciatus* mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

As outlined in the suggested conditions for importation, these mosquitoes will either contain the same wild type bacterium (*Wolbachia* spp.) which is already endemic in *Culex quinquefasciatus* mosquitoes in Hawaii, or will be inoculated with an incompatible bacterium (*Wolbachia* spp.) that is not native to the wild mosquito's current internal fauna. The presence of this different strain of bacteria within the male mosquito's reproductive system will render the imported male mosquitoes unable to successfully mate with wild females found within Hawaii, a process called cytoplasmic incompatibility. Cytoplasmic incompatibility has been used with much success in other parts of the world to reduce mosquito populations and thus reduce the potential of transmission of mosquito vectored diseases. We intend to import male, sexually incompatible mosquitoes for direct release onto the environment. This process uses cytoplasmic incompatibility to reduce current populations of this pest mosquito species, which are vectors for pathogens to Hawaii's fauna, including pathogens such as avian malaria, and which can vector West Nile virus, and lymphatic filariasis to humans. Importing Hawaii lineage mosquitoes which contain the wild type bacterium, will ensure that we can conduct genetic analysis to confirm that the wild *Culex quinquefasciatus* is the wild type originally provided to the collaborators, and that the inoculated mosquitoes are indeed incompatible.

Culex quinquefasciatus is an invasive, disease-spreading mosquito that has dispersed across the Hawaiian islands since its accidental introduction in the 1800s. The species is present on Hawaii, Maui, Molokai, Lanai, Kahoolawe, Oahu, Kauai, and the northwest

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Hawaiian islands. *Culex quinquefasciatus* can thrive at sea-level to 4800ft in elevation. In Hawaii, the mosquito is able to transmit pathogens to native forest birds. The spread of avian malaria, in particular, has contributed to the extinction of more than half of Hawaii's endemic honeycreepers and continues to pose a risk to the remaining species. *Culex quinquefasciatus* is also known to transmit dog heartworm within pets found throughout Hawaii, and is a concern to human health given its ability to vector West Nile virus on the US mainland and lymphatic filariasis in other Pacific nations.

Efforts to suppress *Culex quinquefasciatus* through utilization of traditional vector control methods (e.g., pesticides) are inadequate at a landscape scale, and may be problematic for other non-target state and federally protected invertebrate species including Hawaiian picture-wing flies (*Drosophila* spp.), damselflies (*Megalagrion* spp.), yellow-faced bees (*Hylaeus* spp.) and anchialine pond shrimps (*Vetericaris chaceorum* and *Procaris hawaiiana*). Current efforts to control mosquito-vector disease outbreaks are limited to reducing mosquito breeding site locations and localized applications of various larvicides and adulticides.

On September 6-7, 2016, local, national, and international experts gathered in Hawaii to discuss how to mitigate mosquito-borne diseases. The strategy deemed most favorable in terms of its effectiveness, technical readiness, and safety was *Wolbachia*-based cytoplasmic incompatibility. Cytoplasmic incompatibility results from the presence of a bacterium, *Wolbachia*, in the cells of the mosquito. Many arthropod species, including several native species here in Hawaii, naturally contain strains of *Wolbachia*. Bacteria in the genus *Wolbachia* are a type of arthropod endosymbiont that do not occur in humans or other vertebrates. Approximately 50% of insect species naturally have the bacteria, although many of these insects can survive without *Wolbachia*. Conversely, *Wolbachia* cannot persist outside of insect cells, as it is an obligate endosymbiont. The largest effect of *Wolbachia* is on mating compatibility between individual insects that carry the bacteria. However, there are secondary effects that are being studied by many labs. These include altered host insect lifespan and reduced vector competence.

In nature, *Wolbachia* are passed from females to their offspring. Different strains of *Wolbachia* have also been introduced into insects in laboratories. If a male mosquito with one type of *Wolbachia* mates with a female mosquito that has a different strain of *Wolbachia* the resulting offspring can be inviable and not develop into mosquito larvae because of a mismatch of cellular signals (loss of the male parental chromosomes) originating from *Wolbachia*. If sufficient numbers, on the order to 10 times the wild population size, of male mosquitoes of a different *Wolbachia* type are released, wild females are more likely to mate with males of a different *Wolbachia* type and are predicted to have far fewer viable offspring. With subsequent releases, this process can significantly suppress the wild population numbers of mosquitoes over the following generations over a geographic area. *Wolbachia* male-based insect control programs have been highly successful for reducing local mosquito populations around the world. Results of initial trials in Fresno, California showed decrease of biting *Ae. aegypti* females by 68%, 95%, and 84% during the peak mosquito seasons in 2017, 2018, and

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2019 respectively. *Wolbachia* cannot be spread by the released males, because *Wolbachia* are only passed from mother to offspring. It is also worth noting that male mosquitoes do not bite or vector disease.

One way to generate mosquitoes with a different *Wolbachia* type, is by clearing the naturally-occurring *Wolbachia* strain from the mosquitoes using the antibiotic tetracycline. Then *Wolbachia* can be harvested from cells of another insect species (this can be another mosquito or a non-mosquito species) and introduced into the cleared mosquitoes via microinjection. Another method to establish new *Wolbachia* strains is to mate a *Wolbachia*-carrying female insect to males that have been cleared of their naturally-occurring *Wolbachia* via antibiotic treatment. Because *Wolbachia* are maternally inherited (described above), this cross results in all of the offspring inheriting whichever *Wolbachia* strain is contained in the female parent. Incompatible *Wolbachia* strains can also be naturally present in populations of mosquitoes.

The first shipper listed within this import application, MosquitoMate Inc., holds the US patent, Patent No.: US 7,868,222 B1, for the method of producing an artificial infection in Culicidae species.

<https://patentimages.storage.googleapis.com/55/da/ae/d7cb8b9cb44599/US7868222.pdf>

Additionally, MosquitoMate Inc. offers a commercially available, *Wolbachia* infected male mosquito product for purchase to suppress *Aedes albopictus* mosquito populations via cytoplasmic incompatibility. This product, ZAP Males®, has been reviewed and registered under the Federal Insecticide, Fungicide, and Rodenticide Act (FIFRA). ZAP Males® are a labeled pesticide product with the EPA registration number 89668-4. This product currently has a restriction that only allows for its application in specific states, which does not currently include Hawaii. We reference this, as it is the only registrant in the US currently with a *Wolbachia* mosquito product currently in process of registration with the EPA.

https://www3.epa.gov/pesticides/chem_search/ppls/089668-00004-20171103.pdf

The second shipper listed is Verily Life Sciences, a CA based company which is in the process of working with a different incompatible *Culex quinquefasciatus*. This company is initiating consultations with the EPA relating to this different *Wolbachia* mosquito and will provide additional information directly to HDOA as needed.

Culex quinquefasciatus mosquito eggs originating from Hawaii stock (aka collected from field sites in Hawaii) have been provided to MosquitoMate and Verily for development and testing of cytoplasmic incompatibility. These mosquitoes have been crossed with female mosquitoes carrying a different *Wolbachia* species as outlined above. These mosquitoes have then been backcrossed with a separate population of mosquitoes originating from Hawaii stock over at least seven generations to ensure Hawaii's wild mosquito genetics are >99% contained within a commercially available product to be applied within Hawaii.

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Generations	HI Mosquito Genetics	Crossed MosquitoMate Genetics
0	100.00%	100.00%
1	50.00%	50.00%
2	75.00%	25.00%
3	87.50%	12.50%
4	93.75%	6.25%
5	96.88%	3.13%
6	98.44%	1.56%
7	99.22%	0.78%
8	99.61%	0.39%
9	99.80%	0.20%
10	99.90%	0.10%

On January 17, 2017, the Hawaii Invasive Species Council, an inter-departmental collaboration of the Departments of Land and Natural Resources (DLNR), Agriculture (HDOA), Health (HDOH), Transportation (DOT), Business, Economic Development & Tourism (DBEDT), and the University of Hawaii (UH) passed resolution 17-2, specifically pertaining to mosquitoes. Resolution 17-2, entitled, “Supporting Evaluation and Implementation of Technologies For Landscape-Scale Control of Mosquitoes, With a Focus On Mitigating Both Human and Wildlife Health Risks,” recognizes that mosquitoes in the State of Hawaii are non-native and an important pest species to control. The resolution supports the implementation of evaluated technologies that are scientifically demonstrated as safe, effective control measures for mosquitoes. (<https://dlnr.hawaii.gov/hisc/files/2013/02/HISC-Reso-17-2-signed.pdf>).

House Resolution (HR) 297 passed the Hawaii State House in 2019 and further directed “DOA to review the *Aedes aegypti* mosquito with *Wolbachia* bacteria, including *Aedes aegypti* mosquitoes originating from Hawaii stock that could be imported for landscape scale mosquito control, and render a determination to place it on the appropriate animal import list. Requires DOA, DOH, and DLNR to collaborate on a report to the Legislature with recommendations for appropriate vector control programs.” (https://www.capitol.hawaii.gov/session2019/bills/HB297_SD1_.htm)

Additionally, House Resolution (HR) 95 passed the Hawaii State House in 2021 urging DLNR, DOA, DOH and UH to implement a mosquito control program using *Wolbachia* to reduce mosquito population levels throughout the state. (https://www.capitol.hawaii.gov/session2021/bills/HR95_HD1_.htm)

It should be noted that this project has been developed with the full support of, and will be implemented in close coordination with, the Hawaii Department of Health Vector Control Branch. Per Hawaii Revised Statutes §26-13, the Department of Health “shall administer programs designed to protect, preserve, care for, and improve the physical and mental health of the people of the State.” DOH has the authority in Hawaii relating to mosquitoes and public health, and their staff have decades of expertise to implement

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mosquito surveillance, control and abatement programs.

Proposed Required Conditions for Importation via HDOA List of Restricted Animals Part A

Included are proposed conditions, suggested in collaboration with the HDOH Vector Control Branch, that could be required for importation if *Culex quinquefasciatus* mosquitoes are added to the HDOA List of Restricted Animals Part A to ensure any future imports meet safeguards to preserve public health, the environment, and the long-term efficiency of the IIT tool. All of the following suggested requirements would need to be met to obtain importation permitting.

Culex quinquefasciatus

1. Only mosquitoes originating from a Hawaii stock are allowed for importation.
2. Only mosquitoes containing the same wild-type bacteria as is already present in Hawaii, or a sexually incompatible *Wolbachia* bacteria compared against Hawaii's wild mosquito populations are allowed for importation.
3. Only adult male mosquitoes are allowed for importation.
4. Only individuals or organizations who have conducted work for EPA registration trials for mosquito biopesticide products and who can provide data on rearing and sorting methodologies are allowed to ship these mosquitoes to Hawaii.
5. Only individuals or organizations listed on the import application are allowed to import/receive these mosquitoes.
6. Only islands with established or incipient wild mosquito populations, as determined by the Hawaii Department of Health's Vector Control Branch, are allowed to import these mosquitoes.
7. All environmental review processes, including potential Environmental Impact Statements, Environmental Assessments, or other environmental compliance requirements as outlined by State Law and OEQC, must be completed or cited prior to importation.

Specific details for importation

This is an application for:

- A permit to import male, mosquito species: *Culex quinquefasciatus*.
- The listing of these mosquito species on the Hawaii Department of Agriculture's (HDOA) List of Restricted Animals Part A given that specific conditions, as outlined and enforced by HDOA, are met at the time of importation. Suggested conditions for importation are included within this application.

Within *Culex quinquefasciatus*, the strain of incompatible bacterium will be *Wolbachia wAlbA*, *Wolbachia wAlbB*, or *Wolbachia wPip4*. These *Wolbachia* bacterium are not present within the corresponding species of Hawaii's established mosquito population. The presence of this bacterium will make these males sexually incompatible with the

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wild, established female mosquitoes. Once imported, the male, sexually incompatible males will be released according to EPA and HDOA label directions to suppress the population of the established mosquito populations. Based on the prior use of this technology in California, Florida, and Kentucky, there are no data to suggest releases of these male mosquitoes to have a negative impact on agriculture, the environment, or public health and safety. Existing wild-type bacteria strain that may be imported is wPipV, which is already found on all of the main Hawaiian islands.

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Locations and Safeguards

All mosquitoes for import will originate from Hawaii biotypes collected from Hawaii. All mosquitoes will be backcrossed for at least 7 generations to ensure >99% Hawaii genetics are contained within the commercially available products to be applied within Hawaii. This backcrossing will also mitigate the risks of infections microorganisms and parasites to the mosquitoes via vertical transmission – thus lowering the risk of the mosquitoes accidentally introducing a new parasite or pathogen. In order for these mosquitoes to acquire and vector a disease, an adult female must blood feed from a disease infected vertebrate, and the pathogen must survive in the mosquito and be

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injected into another vertebrate during a subsequent blood feeding. As the intended importation of these mosquitoes only includes the importation of male mosquitoes that do not bite or feed on blood, the unintended importation of an acquired pathogen is eliminated. Verification of Hawaii biotypes and *Wolbachia* strains will be conducted on initial shipments of male mosquitoes to verify requirements have been met, in collaboration with University of Hawaii and Department of Health.

These mosquitoes will be imported into Hawaii through the use of commercial cargo flights. Upon reception to Hawaii, the male mosquitoes will be directly released into the laboratory for quality control testing, and into the environment for the purpose of suppressing the wild mosquito populations. These releases will be performed by individuals or organizations certified to apply these mosquito pesticide products to ensure that the product will be applied properly according to the recommended guidelines.

MosquitoMate and Verily will regularly sample release containers by releasing the contents into lab cages and then examining mosquito sex and number. There is an EPA reviewed value of 1 female release per 250,000 males with the MosquitoMate product. A similar value is likely to be estimated for *Culex quinquefasciatus* given that similar automation, engineering and machine learning technology is being applied to sex sorting. MosquitoMate and Verily have not previously identified a female in a single release container during the course of the Puerto Rico or Fresno projects. In another example, a published study estimates the probability at less than 1 female per 200 million males (Crawford JE, Clarke DW, Criswell V, Desnoyer M, Cornel D, Deegan B, et al. Efficient production of male *Wolbachia*-infected *Aedes aegypti* mosquitoes enables large-scale suppression of wild populations. *Nat Biotechnol.* 2020;38(4):482-92.) To date, PCR monitoring of mosquitoes collected from release field sites have not identified any ZAP infected females.

At least once per year, MosquitoMate and Verily will also conduct longevity and competitiveness studies, comparing the mosquitoes proposed for releases and wild type males. Data from previous trials demonstrate ZAP mosquito longevity and competitiveness to be at least equal to Wild Type males. In addition to Hawaii's import requirements, the shipper and/or receiver will obtain additional permits as required by federal or state agencies.

Wolbachia is an obligate endosymbiont and cannot survive outside of the host invertebrate. *Wolbachia* strains already exist in Hawaii in a range of invertebrates in the wild, including mosquitoes. The presence of *Wolbachia* endosymbionts is the normal

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state for 40% to 60% of Arthropods and does not represent an unusual or pathogenic bacterial infection. *Wolbachia* are not capable of infecting human cells. MosquitoMate and Verily will perform PCR testing on the mosquitoes to confirm the presence of the correct *Wolbachia* bacterium within the shipment lineage to ensure cytoplasmic incompatibility.

The likelihood that introduced strains of *Wolbachia* would become the dominant strains in the environment is highly unlikely. Replacing the dominant *Wolbachia* strain has been done purposefully in the environment for projects that are separate from the approach we are proposing (such as by the World Mosquito Program in Australia and other nations). To clarify, DLNR is NOT proposing a World Mosquito Program type project where the goal is to intentionally force a different dominant *Wolbachia* strain into the wild mosquitoes in the environment and change vector competence of the wild population. However, in these types of programs, they have to release 4 million mixed male AND female mosquitoes in a given location to force a new *Wolbachia* strain to become the dominant strain over an area of 66 km². Given the aforementioned EPA reviewed value of 1 female release per 250,000 males with the MosquitoMate product, such an outcome is not expected to occur.

If, somehow population replacement were to occur (despite the estimated 1 female release per 250,000 males) DLNR would cease releases as the released males would then be able to mate with the wild females with the established *Wolbachia* species. The outcome of this would be that the mosquito species that already exists in Hawaii would continue to exist in the wild, just with a different *Wolbachia* bacteria. We do not anticipate a different *Wolbachia* bacteria having any new or negative effects on the environment.

DLNR and DOH feel comfortable utilizing these mosquitoes at a very small scale (in remote forest habitat) or at a very large scale (across urban areas and island wide) so long as recommended application guidelines are followed. The scale and scope of the project will likely vary across time based on the funding available and mosquito prevalence. As with any pesticide product, if you do not eradicate the species of concern, they will rebound if you stop using the pesticide product. However, we view this as a beneficial aspect of the project as we also know we can stop the process at any time. Unfortunately, due to the critical nature of the declines of Hawaiian forest birds, we anticipate mosquito control becoming a long-term management action to be performed (similar to rat control and invasive weed control) annually.

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Data collection will occur during releases using the State general funds as well as federal funds from partner agencies (USFWS, USGS, NPS), depending on who is performing the releases. As the application of the pesticide product is intended for the reduction of *Culex quinquefasciatus* mosquito populations, this monitoring will include extensive mosquito population surveillance following releases to ensure that populations are reduced. DLNR is already conducting this type of monitoring in preparation for incompatible mosquito releases. *Wolbachia* genetic monitoring will also occur, likely in partnership with USGS, throughout the release program.

In addition to Hawaii's import requirements, the shipper and/or receiver will obtain additional permits as required by federal or state agencies.

Method of Disposition

Any dead imported mosquitoes will be disposed of as municipal waste.

Abstraction of Organism

Culicidae species are sexually reproducing species. Minimum generation times vary but are approximately three weeks. Mature adults are up to approximately a centimeter in length and can live for a month to a few months. Adult mosquitoes range from 2.0 to 10.0 mm in size with males being smaller than females on average. Mosquito life cycles are well understood for most species, including all those established in Hawaii.

Larvae feed on organic material found in pools of water. Both adult males and females feed on water that contains carbohydrates (water with sap or nectar). Only mature females of certain species seek out and feed on vertebrate blood prior to egg laying. This blood feeding process allows for the transmission of pathogens and parasites.

Culex quinquefasciatus rely on pools of water with organic material for the growth of larvae. Only adult females bite, as they require blood meals from vertebrate hosts to develop their eggs.

Potential Impact to the Environment

Culex quinquefasciatus are already well established in the wild on all of the main islands in Hawaii from sea-level to ~6,000 feet in elevation. and *Culex quinquefasciatus* are established statewide and is well establish on Hawaii's Big Island. An additional five other "biting" non-native mosquito species have also become established: *Ae. albopictus*, *Ae. aegypti*, *Ae. japonicus*, *Ae. vexans*, and *Wyeomyia mitchelli*.

Wolbachia are not infectious to humans and are vertically transmitted through the eggs

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from one generation to another. The *Wolbachia* bacteria are obligate endosymbionts and can only survive inside the insect host's cytoplasm. A mosquito transinfected with a different strain of *Wolbachia* that results in cytoplasmic incompatibility would not be able to successfully reproduce with a wild mosquito due to cytoplasmic incompatibility.

Therefore, if individual mosquitoes did become temporarily established, then they will quickly die off over the following generations because of cytoplasmic incompatibility with wild mosquitoes of the same species, with which they would be expected to encounter and mate.

Through the importation we intend to only import male mosquitoes. The sex separation can be performed in a variety of manners including through computer recognition and separation of males and females or through pupal sorting of males and females.

However, if both sexes of transinfected mosquito were to be accidentally released, they are unlikely to maintain a breeding population of a transinfected mosquito. *Wolbachia* invasions into populations require a critical threshold frequency of infection that needs to be overcome before a novel *Wolbachia* infection can spread into a population. The *Wolbachia* infection rate must exceed 20-45% before it can spread and become established. This is evident in large scale releases such as in Cairns, Australia, where millions of transinfected mosquitoes (both sexes) with *Wolbachia* are released into the environment to control disease transmission, yet they do not easily reach fixation in the wild. If transinfected mosquitoes were to become established, the establishment is likely to be spatially localized due to incompatibility with neighboring mosquito populations.

Potential Impacts of Importation

pro: Importation of male mosquitoes will allow the implementation of an evaluated technology that has been scientifically demonstrated as a safe and effective control method for mosquitoes on a landscape-scale. These are mosquitoes that are widespread in Hawaii and which have negative impacts to humans, wildlife, and pets, and are causing the extinction of native forest birds. Thirty species of main Hawaiian forest birds have become extinct since European contact, and another 11 of the 21 remaining species are federally listed as threatened or endangered. The remaining 21 forest bird species remain at great risk as a result of avian pox and avian malaria. Four honeycreeper species (*Akikiki*, *Oreomystis bairdi*; *Akekee*, *Loxops caeruleirostris*; *Kiwikiu*, *Pseudonestor xanthophrys* and *Akohekohe*, *Palmeria dolei*) are of particular concern – each are federally endangered, single-island endemics with highly restricted ranges, number fewer than 1,800 individuals, and display recent alarming population declines. DLNR and USFWS have previously attempted to address these declines through bold conservation actions, such as translocations and establishment of captive

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populations; however agencies have met with only limited success due to rapidly changing disease-transmission conditions on the landscape. There is an urgent need to develop new conservation tools, including landscape-level mosquito control in order to prevent further extinctions.

The application of traditional chemical controls for mosquitoes in both natural areas is impractical and causes unacceptable non-target impacts, whereas IIT carries no non-target risks to native species, humans or the environment. Furthermore, mosquitoes were first introduced to the Hawaiian Islands in the 1800s, and while they are used opportunistically as prey items, no species native to Hawai'i are dependent on their presence for survival. The control of mosquito populations in Hawaiian forests would thus cause no negative impacts on Hawaiian species.

Demonstrated application of this approach in Hawaii would also have a wide range of potential positive effects in that it may facilitate the incompatible insect technique approach being used for human health.

con: It is hard to imagine any negative effects since the species is already established in Hawaii. Importing these organisms will not have any foreseeable beneficial effect to this mosquito species already found in Hawaii. The introduction of, for example, increased genetic variation within the mosquito species will be minimized by crossing the lines to mosquitoes originating from Hawaii.

The presence of unintended accompanying microbiota is minimized by the sterile laboratory rearing conditions used. These mosquitoes have been maintained for many generations in the lab environment and have not had the opportunity to obtain pathogens from the wild from blood feeding. The presence of intended microbiota, the *Wolbachia*, potentially has very positive effects on the societal health, the suppression of human disease vectored by mosquitoes, the environment, via population suppression of mosquitoes that vector avian pathogens, and the economy, through the potential increased tourism and lessened disease burden.

This mosquito species is already well established in Hawaii, as are many different strains of *Wolbachia*. MosquitoMate and Verily have a demonstrated track record of success utilizing sex-sorting methods which are highly effective. In the event that technical difficulties did occur during sex-sorting methods, because of cytoplasmic incompatibility, the escape of female mosquitoes carrying a new *Wolbachia* strain is not expected to be stable over the following generations. Laboratory reared females outcrossing to locally established wild male mosquitoes will result in cytoplasmic

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incompatibility and the failure of offspring to develop.

There is an extensive body of literature surrounding this mosquito species, its impact upon Hawaii, and Wolbachia-mediated cytoplasmic incompatibility.

Information on *Wolbachia*, with a focus on cytoplasmic incompatibility within mosquitoes:

- Atyame, C. M., Cattell, J., Lebon, C., Flores, O., Dehecq, J.-S., Weill, M., Gouagna, L. C. & Tortosa, P. (2015) Wolbachia-based population control strategy targeting *Culex quinquefasciatus* mosquitoes proves efficient under semi-field conditions. *PLoS ONE* 10, e0119288.
- Atyame, C. M., Labbé, P., Lebon, C., Weill, M., Moretti, R., Marini, F., Gouagna, L.C., Calvitti, M. & Tortosa, P. (2016) Comparison of irradiation and Wolbachia based approaches for sterile-male strategies targeting *Aedes albopictus*. *PLoS ONE* 11, e0146834.
- Barton, N. H., & Turelli, M. (2011). Spatial waves of advance with bistable dynamics: cytoplasmic and genetic analogues of Allee effects. *The American Naturalist*, 178(3), E48-E75.
- Blagrove, M. S., Arias-Goeta, C., Failloux, A. B., & Sinkins, S. P. (2012). Wolbachia strain wMel induces cytoplasmic incompatibility and blocks dengue transmission in *Aedes albopictus*. *Proceedings of the National Academy of Sciences*, 109(1), 255-260.
- Callaini, G., Dallai, R., & Riparbelli, M. G. (1997). Wolbachia-induced delay of paternal chromatin condensation does not prevent maternal chromosomes from entering anaphase in incompatible crosses of *Drosophila simulans*. *Journal of Cell Science*, 110(2), 271-280.
- Dobson, S. L., Marsland, E. J., & Rattanadechakul, W. (2001). Wolbachia-induced cytoplasmic incompatibility in single- and superinfected *Aedes albopictus* (Diptera: Culicidae). *Journal of Medical Entomology*, 38(3), 382-387.
- Hamm, C. A., Begun, D. J., Vo, A., Smith, C. C., Saelao, P., Shaver, A. O., ... & Turelli, M. (2014). Wolbachia do not live by reproductive manipulation alone: infection polymorphism in *Drosophila sukuzii* and *D. subpulchrella*. *Molecular Ecology*, 23(19), 4871-4885.
- Hoffmann, A. A., Montgomery, B. L., Popovici, J., Iturbe-Ormaetxe, I., Johnson, P.H., Muzzi, F., ... & Cook, H. (2011). Successful establishment of Wolbachia in *Aedes* populations to suppress dengue transmission. *Nature*, 476(7361), 454.
- Hoffmann, A. A., Ross, P. A., and Rašić, G. (2015) Wolbachia strains for disease control: ecological and evolutionary considerations. *Evolutionary*

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Applications, 8(8),751-768.

Jiggins FM. (2017) The spread of *Wolbachia* through mosquito populations.

PLoS Biology 15(6):e2002780.

Laven, H. (1967). Eradication of *Culex pipiens fatigans* through cytoplasmic incompatibility. *Nature*, 216(5113), 383-384.

Liao W, Atkinson CT, LaPointe DA, Samuel MD (2017) Mitigating Future Avian Malaria Threats to Hawaiian Forest Birds from Climate Change. *PLoS ONE* 12(1):e0168880.

Mains, J. W., Brelsfoard, C. L., Rose, R. I. & Dobson, S. L. (2016) Female adult *Aedes albopictus* suppression by *Wolbachia*-infected male mosquitoes. *Scientific Reports* 6, 33846.

Stouthamer, R., Breeuwer, J. A., & Hurst, G. D. (1999). *Wolbachia pipientis*: microbial manipulator of arthropod reproduction. *Annual Reviews in Microbiology*, 53(1), 71-102.

Sinkins, S. P., Braig, H. R., & O'Neill, S. L. (1995). *Wolbachia* superinfections and the expression of cytoplasmic incompatibility. *Proceedings of the Royal Society of London B: Biological Sciences*, 261(1362), 325-330.

Tram, U., & Sullivan, W. (2002). Role of delayed nuclear envelope breakdown and mitosis in *Wolbachia*-induced cytoplasmic incompatibility. *Science*, 296(5570), 1124-1126.

Waltz, E. (2016) US reviews plan to infect mosquitoes with bacteria to stop disease. *Nature* 533, 450-451.

Weinert, L. A., Araujo-Jnr, E. V., Ahmed, M. Z. & Welch, J. J. (2015) The incidence of bacterial endosymbionts in terrestrial arthropods. *Proc. R. Soc. B* 282, 20150249.

Werren, J. H., Baldo, L., & Clark, M. E. (2008). *Wolbachia*: master manipulators of invertebrate biology. *Nature Reviews Microbiology*, 6(10), 741.

Zug, R., & Hammerstein, P. (2012). Still a host of hosts for *Wolbachia*: analysis of recent data suggests that 40% of terrestrial arthropod species are infected. *PLoS ONE*, 7(6), e38544.

***Wolbachia* in Hawaii:**

Atkinson, C. T., W. Watcher-Weatherwax, and D. A. LaPointe. (2016) Genetic diversity of *Wolbachia* endosymbionts in *Culex quinquefasciatus* from Hawaii, Midway Atoll and American Samoa. Technical Report HCSU-074.

Bennett, G. M., Pantoja, N. A., & O'Grady, P. M. (2012). Diversity and phylogenetic relationships of *Wolbachia* in *Drosophila* and other native Hawaiian insects. *Fly*, 6(4), 273-283.

Other References:

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Anonymous. (2017). To Restore a Mosquito-Free Hawaii. Summary Report of the Workshop to Formulate Strategic Solutions for a “Mosquito-Free Hawaii”. Available at: “http://www.cpc-foundation.org/uploads/7/6/2/6/76260637/report_on_mosquito_free_workshop.pdf.”

Anonymous. (2020). Three great years of Debug Fresno. Available at: “<https://blog.debug.com/2020/01/three-great-years-of-debug-fresno.html>”

Dame, D. A., Curtis, C. F., Benedict, M. Q., Robinson, A. S., & Knols, B. G. (2009). Historical applications of induced sterilisation in field populations of mosquitoes. *Malaria Journal*, 8(2), S2.

Johnston, D., *et al.* (2016). Notes from the field: outbreak of locally acquired cases of dengue fever—Hawaii, 2015. *MMWR. Morbidity and Mortality Weekly Report*, 65(2); 34–35.

Kauffman, E., Payne, A., Franke, M. A., Schmid, M. A., Harris, E., & Kramer, L. D. (2017). Rearing of *Culex* spp. and *Aedes* spp. mosquitoes. *Bio-Protocol*, 7(17).

Mendenhall, I. H., S. A. Tello, L. A. Neira, L. F. Castillo, C. B. Ocampo, and D. M. Wesson. (2012) Host Preference of the Arbovirus Vector *Culex erraticus* (Diptera: Culicidae) at Sonso Lake, Cauca Valley Department, Colombia. *Journal of Medical Entomology* 49 (5): 1092–1102

O'Neill, S. L.; Ryan, P. A.; Turley, A. P.; Wilson, G.; Retzki, K.; Iturbe-Ormaetxe, I.; Dong, Y.; Kenny, N.; Paton, C. J. & Ritchie, S. A. (2018) Scaled deployment of *Wolbachia* to protect the community from *Aedes* transmitted arboviruses. *Gates Open Research* 2: 1-18

Richmond JY, McKinney RW, eds. (1999). *Biosafety in Microbiological and Biomedical Laboratories* (4th ed.). ISBN 0-7881-8513-6.

Scott, T. W. (2005). Containment of arthropod disease vectors. *ILAR Journal*, 46(1), 53-61.

Takken, W., & Verhulst, N. O. (2013). Host preferences of blood-feeding mosquitoes. *Annual Review of Entomology*, 58, 433-453.

[USFWS] U.S. Fish and Wildlife Service. (2006) Endangered and Threatened Wildlife and Plants; Determination of Status for 12 Species of Picture-Wing Flies from the Hawaiian Islands. *Federal Register* 71 (89): 26835-26852.

Winchester, J. & Kapan, D. (2013). History of *Aedes* Mosquitoes in Hawaii. *Journal of the American Mosquito Control Association* 29(2): 154-163.

Questions from Hawaii Department of Agriculture on Incompatible Culex strain

The following information is being provided by Verily Life Sciences in response to questions from the Hawaiian Department of Agriculture (HI-DoA) about the *wAlbB*-strain *Culex quinquefasciatus* (incompatible i.e. conditionally sterile mosquitoes), which the US Fish and Wildlife Service, the Birds Not Mosquitoes (BNM) coalition and other collaborators wish to import into Hawaii for use in a mosquito control program that uses Sterile Insect Technique (or in this case, an incompatible insect technique). The objective of this program is to protect native Hawaiian birds against avian malaria, which is vectored by invasive *Cx quinquefasciatus* in Hawaiian bird reserves.

Background information:

Debug is a Verily Life Sciences (Verily) project aimed at developing technology to rear and release sterile or incompatible mosquitoes to reduce mosquito populations that transmit disease. Much of the Debug project's work has focused on developing tools for the mass rearing and effective release of male mosquitoes that are conditionally sterile against Wild Type (WT) mosquitoes due to cytoplasmic incompatibility (CI): whereby male insects infected with *Wolbachia* that mate with either females without *Wolbachia* or those infected with different *Wolbachia* strain, produce non-viable progeny, as embryonic development is halted. CI is the basis for the *Wolbachia* Sterile Insect Technique (SIT), also referred to as the Incompatible Insect Technique (IIT). Debug has successfully developed and deployed incompatible male *Aedes aegypti* mosquitoes in several large scale collaborative programs. For reference the following publications provide detail about the efficacy, general approach, rearing, sex sorting, release and field results of some of these projects:

- [Crawford et al 2020](#), Fresno CA, USA (2017-2018)
- [Beebe et al 2021](#), Innisfail QLD, Australia (2018), see also [Australian CSIRO website on the Innisfail project](#).
- [Ng et al, 2021](#), Singapore (2018-current), see also [Singapore National Environment Agency website on Project Wolbachia Singapore](#)

While there are no recent publications on field programs using incompatible *Cx. qunig.*, there is a 1967 publication showing local elimination of *Culex pipiens* using release of incompatible *Wolbachia* males in Myanmar (Laven, H. Eradication of *Culex pipiens fatigans* through Cytoplasmic Incompatibility. *Nature* 216, 383–384 (1967). <https://doi.org/10.1038/216383a0>.), and incompatibility in *Cx. pipiens* is widely studied.

In Hawaii, Debug has been requested by Hawaiian conservation groups including the Birds Not Mosquitoes coalition, The Nature Conservancy and the US Fish and Wildlife Service, to participate in a project that attempts to protect native Hawaiian birds from the depredations of Avian Malaria vectored by the invasive *Cx. qunig.* mosquitoes. This project would release

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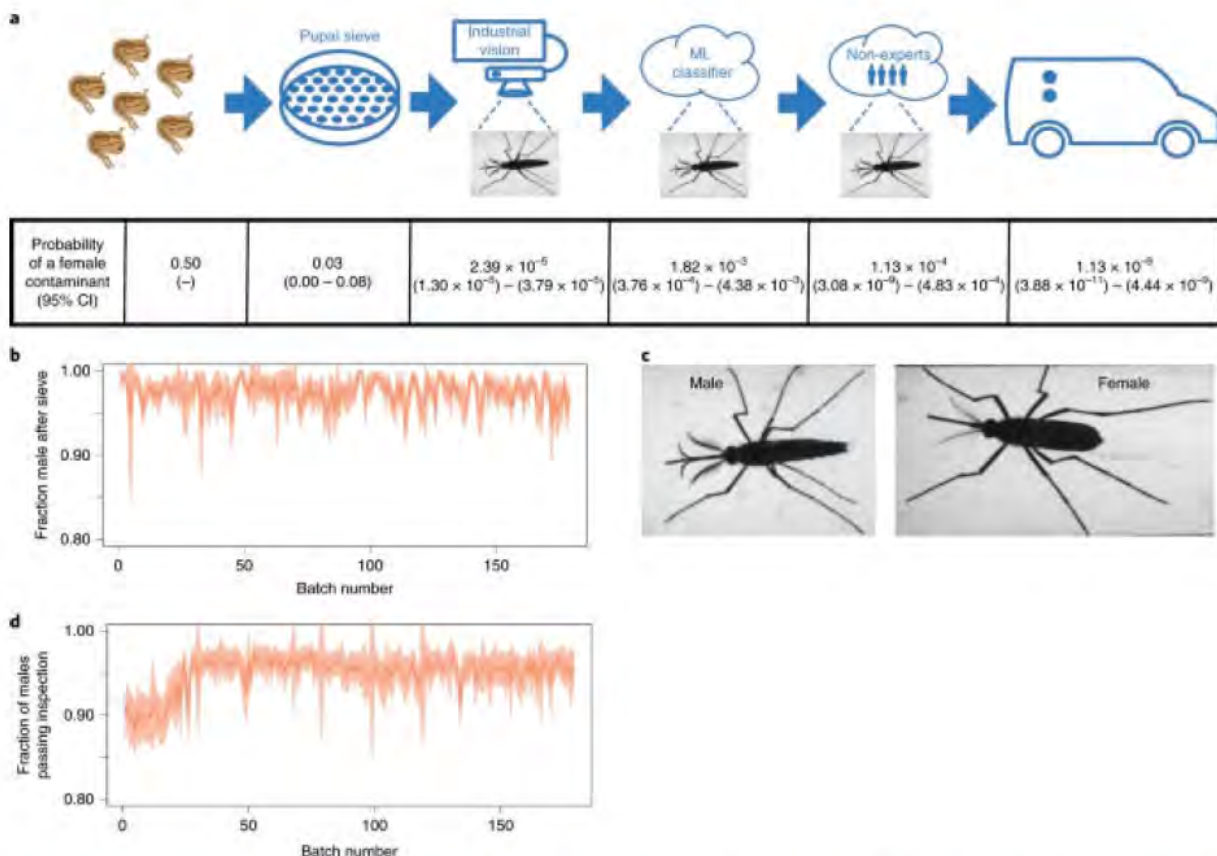
incompatible male *Cx. quinq.* into forest reserves with the goal of reducing wild-type (WT) *Cx. qunig.* mosquitoes and hence reducing malaria infections in birds. These male mosquitoes have a short lifespan (days), can not bite or vector disease, and are conditionally sterile due to the presence of an incompatible strain of the endosymbiont *Wolbachia pipientis* already present in Hawaiian mosquitoes (*wAlbB* sourced from Hawaii.) As noted previously *Cx. quinquefasciatus* are widespread in Hawaii, *Wolbachia pipientis* is present in a majority of insect species, including many endemic to Hawaii, all *Culex* and *Aedes albopictus* mosquitoes in HI have *Wolbachia pipientis*, and thus there is a long history of human and ecological exposure.

Verily's understanding is that the Hawaiian Department of Agriculture intends to submit an emergency application under FIFRA Section 18 to the EPA for temporary registration of the *wAlbB* in *Cx. quinq.* and that releases will be conducted under appropriate permits.

Questions from Christopher Kishimoto (Hawaii Department of Agriculture, Plant Quarantine Branch, Entomologist)

1. Could you please walk us through your procedures on how you eliminate female mosquitos from being released?

Verily's multi-stage mosquito sex sorting system removes females with extremely high accuracy, while retaining most males for release. As outlined in [Crawford et al. 2020](#) the system comprises 3 stages: 1) sieving of pupae to remove the vast majority of females, as female pupae are larger than males, 2) adult sex sorting using a real-time industrial vision system that uses multiple images of every adult mosquito to identify and remove females, and finally 3) we submit all images of individuals labeled male by the industrial vision system for scoring by a machine learning classifier which acts as a quality control system with further human review to identify and enable removal of any potential females accepted by the adult industrial vision sorter. For *Aedes aegypti* the combined system is expected to release 1 female for every 900 million males with a 95% CI of 1:200 million to 1:26 billion ([Crawford et al. 2020](#)). We are currently adapting the Verily sex sorting pipeline to *Culex* mosquitoes but as the basic features used for sex sorting, particularly the adult morphology, are the similar between *Culex* and *Aedes* we hope for extremely high accuracy sorting. As we do for *Aedes* as a part of EPA permitted manufacturing, we will also perform regular QC assays to confirm absence of females in sample release batches prior to the beginning of shipments and importation into Hawaii. See diagram below from [Crawford et al. 2020](#)



a, Illustration of the entire [*Aedes aegypti* mosquito] sex-sorting pipeline, including the mechanical pupal sieve, real-time adult visual inspection, cloud-based machine learning classifier, and [expert human] review. The probability of a female contaminant with 95% CIs for each step is shown along with the estimated overall female contamination rate for the entire pipeline in the final column. [note: the sorting process has been updated since 2018 to include both expert review and other algorithmic improvements] b, The fraction of mosquitoes imaged by the sex sorter after the pupal sieve that were male with s.d. intervals shaded for 179 production batches. c, Example images from the adult sex sorter (male on the left and female on the right) used by both the industrial vision system and machine learning classifier. d, The fraction of true males that were correctly labeled and accepted by the Industrial Vision system with s.d. interval shaded (n=mean of 96, range of 10–140 independent sex-sorter lane measurements per batch). After the industrial vision stage there are further QC inspection steps as noted above.

2. How often are quality control measures implemented?

Every single mosquito Verily provides for release goes through the above sex-sorting pipeline, with multiple stages of independent computer review of each adult mosquito, followed by additional human and computer quality control reviews.

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Sieve performance is monitored in every single batch, and in addition as a part of the documented (and EPA reviewed) manufacturing process Verily regularly conducts a “female contamination” assay to ensure that adult sorting runs let through no females, validating that release batches are at <1:250:000 females:males according to Verily’s calculations and EPA requirements.

3. How often will quality control checks be implemented in the future?

As noted above, production of *Cx. quinq.* would be under a similar EPA reviewed manufacturing process as used for Verily’s *Aedes* manufacturing. Verily is regularly updating sex sorting algorithms and protocols to increase accuracy.

4. Have female mosquitos ever been found in batches of mosquitos destined for field release? If so, how often?

5. Have batches of mosquitos been halted for distribution because of the findings of any female mosquitos or other problems?

6. Have Verily female mosquitos ever been collected from the environment?

7. How often does Verily survey release sites for Verily female mosquitos?

We will provide answers to questions (4-7) together as they are all aimed at identifying the likelihood of *wAlbB* female releases and identification of this in field environments, which could potentially reduce the efficacy of incompatible male releases.

As a part of Verily’s *Aedes* manufacturing process as documented in Crawford et al. 2020, a small number of females (~<1:250k) are found in batches *prior* to field release. As noted above a secondary quality control review identifies and removes these resulting in extremely low female contamination rate in released mosquitoes. In our Singapore collaboration we identified a very small number of released females in batches comprising several million males, and have since updated our protocols to reduce the likelihood of this recurring.

Regarding the environmental collection of Verily female mosquitoes: The incompatible *Aedes* programs undertaken by Verily and collaborators have Verily (or our partners) routinely monitor trap collections for incompatible adult females i.e. females positive for the released strain of *Wolbachia* as assessed by molecular assays, and we also test larvae from ovitraps for the presence of this *Wolbachia*. We propose that this be included as a part of any fieldwork and surveillance accompanying a release program of incompatible *Culex quinq.* males in Hawaii. We also note that *Wolbachia pipientis* is present in a majority of insect species, including many endemic to Hawaii and all *Culex* and *Aedes albopictus* mosquitoes in HI have *Wolbachia pipientis*, and thus there is a long history of human and ecological exposure.

ZAP (transfected *Ae. albopictus*) and WB1 are MosquitoMate products, and we are unaware of any Verily manufactured ZAP or WB1 females being discovered in the environment. As noted above (and as reported in Ng et al 2021) Verily discovered the accidental release along with field collection of a very small number of *Ae. aegypti* females in Singapore, which despite ongoing release programs have not spread. As noted in the Singapore paper, in response protocols for sex sorting and QC have been significantly improved.

Exact protocols (e.g. sampling rate, pooling, etc.) for field surveillance to identify incompatible *Culex* adult females and/or larvae in Hawaii will need to be agreed and finalized.

8. What are the overall results from field releases of Verily mosquitos so far?

As outlined above in the introduction there are several studies showing that incompatible (transfected) male *Ae. aegypti* can cause suppression of wild mosquito populations when operated using Verily's rearing and release systems.

- California (in collaboration with MosquitoMate and others) showed 95% (up to 99%) suppression in treatment areas relative to controls: Crawford, J.E., Clarke, D.W., Criswell, V. et al. Efficient production of male *Wolbachia*-infected *Aedes aegypti* mosquitoes enables large-scale suppression of wild populations. *Nat Biotechnol* 38, 482–492 (2020). <https://doi.org/10.1038/s41587-020-0471-x>
- Australia (in collaboration with the Australian CSIRO and others) showed >80% (up to 97%) suppression in treatment areas relative to controls: NW Beebe, D Pagendam, BJ Trewin, A Boomer, M Bradford, A Ford, et al. Releasing incompatible males drives strong suppression across populations of wild and *Wolbachia*-carrying *Aedes aegypti* in Australia. *Proceedings of the National Academy of Sciences* 118 (41). <https://www.pnas.org/doi/full/10.1073/pnas.2106828118>
- Singapore [Ng. et al 2021 MedRxiv Preprint Paper](#) in collaboration with Singapore National Environment Agency showed 98% suppression in treatment areas relative to controls.

In addition to recent *Aedes* field results there is a 1967 *Culex pipiens* paper showing local elimination (100% suppression) using release of incompatible males in Myanmar: Laven, H. Eradication of *Culex pipiens fatigans* through Cytoplasmic Incompatibility. *Nature* 216, 383–384 (1967). <https://doi.org/10.1038/216383a0>.

9. How long does it take for wild mosquito populations to get back to prerelease populations once Verily mosquitos have stopped being released?

It is unknown what would happen with *Cx. quinq.* in the proposed program areas after a successful suppression program, as it will depend on migration rates from outside the treatment area, natural fecundity of wild mosquitoes in the local ecology and a variety of other factors. Any release program would need to maintain an ongoing surveillance program to monitor this.

10. Does Verily have EPA approval to release its mosquitos in Hawaii?

11. Does Verily have Hawaii Department of Agriculture Pesticides Branch approval to release its mosquitos in Hawaii?

In answer to 10-11: As previously discussed with HI-DoA, Verily will support partners and HI-DoA in applying for EPA permits along with any state permits required to undertake this program.

12. How many Verily mosquitos would have to be released to achieve adequate population control in Hawaii's environments?

This will be determined by Verily and local partners based on the results of an initial Mark Release Recapture (MRR) trial, which would give information on wild-type population numbers

Verily Life Sciences LLC Response to questions from HI Department of Agriculture

and the dispersion and survival of released incompatible males. In general most IIT programs aim to achieve a ratio of 1:10 Wild Type male:sterile male in field traps in the treatment area to ensure strong suppression in each generation.

13. What is the duration of time needed to achieve adequate mosquito population control once releases start?

This will depend on a number of factors including wild type population, release numbers, efficacy of dispersion etc, along with the efficacy of the surveillance program used to measure impact. *Aedes* incompatible male release programs and other SIT projects typically see measurable impact on hatch-rate within several weeks, though it may take months for significant wild-type population reduction. Laven 1967 saw initial reductions after several weeks with incompatible male *Cx. pipiens* releases though this was in a village setting.

14. How will Verily handle a request to specifically manufacture Hawaiian biotype mosquitos, especially if orders for those mosquitos may be inconsistent?

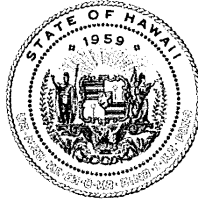
Verily will review with requesting partners as project plans develop.

15. Would Verily be able to show verifiable proof that only Hawaiian biotype mosquitos will be shipped to Hawaii?

Yes, Verily's manufacturing process maintains molecular assays and physical containment to ensure quality controls and biosecurity of shipped mosquitoes.

Sex-sorted male mosquitoes will be shipped to Hawaii from our rearing facility in California and produced using Verily's mosquito manufacturing process which will be reviewed as a part of a HI DoA submitted Section 18 permit application. Males will be transported in line with any issued label and permits.

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ENGINEERING
FORESTRY AND WILDLIFE
HISTORIC PRESERVATION
KAHOOLAWE ISLAND RESERVE COMMISSION
LAND
STATE PARKS

EXEMPTION NOTICE

Regarding the preparation of an environmental assessment under the authority of
Chapter 343, HRS and Section 11-200.1-17, HAR

Project Title:	Mosquito Control Research Using <i>Wolbachia</i> -based Incompatible Insect Technique
Project Location:	<p>Maui</p> <ul style="list-style-type: none"> (2) 2-3-005:004: Waikamoi Preserve (2) 2-4-016:004: Waikamoi Preserve (2) 1-2-004:013: Hanawi Natural Area Reserve (2) 2-3-005:001: Haleakala National Park (2) 1-8-001:007: Haleakala National Park (2) 1-3-001:003: Haleakala National Park (2) 1-7-004:016: Haleakala National Park (2) 1-6-001:001: Haleakala National Park (2) 1-6-001:002: Haleakala National Park (2) 1-2-010:001: Haleakala National Park <p>Kauai</p> <ul style="list-style-type: none"> (4) 1-4-001:003: Alakai Wilderness Preserve (4) 1-4-001:013: Kokee State Park
Chapter 343 Trigger(s):	Use of State Funds and Lands
Project Description:	<p>The main objective of this project is to initiate research to inform incompatible insect technique applications for the control of invasive <i>Culex quinquefasciatus</i> mosquitoes which are the primary vector of avian malaria. The disease threatens the survival of remaining endangered forest bird species where they persist in high elevation montane forest habitat on Maui and Kauai.</p> <p>Male mosquitoes which have been given an incompatible strain of <i>Wolbachia</i> bacteria are to be released on the landscape, and upon release those males will breed with wild female mosquitoes. As a result of those pairings, the wild female mosquitoes will lay eggs which will not hatch, and no offspring will be produced. When releases of incompatible male mosquitoes are completed consecutively, the approach results in the suppression of mosquito populations at a landscape-scale. If releases are halted, mosquito</p>

	<p>populations will gradually return to pre-release levels as wild female and male mosquitoes migrate back into the treated area from surrounding forest habitat. Initial research will contribute to EPA registration of male <i>Culex quinquefasciatus</i> mosquitoes with <i>Wolbachia</i> as a biopesticide, as well as determine the minimum number of male mosquitoes that must be released in each area to ensure population suppression.</p> <p>This project may be funded by Federal sources.</p>
Consulted Parties:	U.S. Fish and Wildlife Service
Authorization:	November 13, 2015, Land Board submittal (C-6). Delegation of Authority to the Chairperson or their authorized representative to declare exempt from the preparation of an Environmental Assessment those Department actions which are included in the Department-wide exemption list when the Board of Land and Natural Resources has delegated the authority to conduct those actions.
Exemption Class & Description:	<p>Exemption Classes:</p> <p>General Exemption Type 5 <i>Basic data collection, research, experimental management, and resource and infrastructure testing and evaluation activities that do not result in a serious or major disturbance to an environmental resource.</i></p> <p>PART 1</p> <p>13. Research that the Department declares is designed specifically to monitor, conserve, or enhance native species or native species' habitat. 16. Research to identify, monitor, control, or eradicate introduced species.</p> <p>Date of Agency Exemption List: November 10, 2020.</p>
Determination:	The Department of Land and Natural Resources declares that this project will likely have minimal or no significant impact on the environment and is therefore exempt from the preparation of an environmental assessment under the above exemption classes.

DES

Suzanne D. Case

Jun 17, 2022

Suzanne D. Case, Chairperson
Board of Land and Natural Resources

Date

Signature:



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Email: david.g.smith@hawaii.gov